



PEARL

Hospital sink traps as a potential source of the emerging multidrug-resistant pathogen *Cupriavidus pauculus*: characterization and draft genome sequence of strain MF1

Butler, James; Kelly, Sean D.; Muddiman, Katie J.; Besinis, Alexandros; Upton, Mathew

Published in:

Journal of Medical Microbiology

DOI:

[10.1099/jmm.0.001501](https://doi.org/10.1099/jmm.0.001501)

Publication date:

2022

Link:

[Link to publication in PEARL](#)

Citation for published version (APA):

Butler, J., Kelly, S. D., Muddiman, K. J., Besinis, A., & Upton, M. (2022). Hospital sink traps as a potential source of the emerging multidrug-resistant pathogen *Cupriavidus pauculus*: characterization and draft genome sequence of strain MF1. *Journal of Medical Microbiology*, 71(2). <https://doi.org/10.1099/jmm.0.001501>

Hospital sink traps as a potential source of the emerging multidrug-resistant pathogen *Cupriavidus pauculus*: characterization and draft genome sequence of strain MF1

James Butler^{1,*}, Sean D. Kelly², Katie J. Muddiman², Alexandros Besinis^{1,3} and Mathew Upton²

Abstract

Introduction. *Cupriavidus pauculus* is historically found in soil and water but has more recently been reported to cause human infection and death. Hospital sink traps can serve as a niche for bacterial persistence and a platform for horizontal gene transfer, with evidence of dissemination of pathogens in hospital plumbing systems driving nosocomial infection.

Gap Statement. This paper presents the first *C. pauculus* strain isolated from a hospital sink trap. There are only six genome assemblies available on NCBI for *C. pauculus*; two of these are PacBio/Illumina hybrids. This paper presents the first ONT/Illumina hybrid assembly, with five contigs. The other assemblies available consist of 37, 38, 111 and 227 contigs. This paper also presents data on biofilm formation and lethal dose in *Galleria mellonella*; there is little published information describing these aspects of virulence.

Aim. The aims were to identify the isolate found in a hospital sink trap, characterize its genome, and assess whether it could pose a risk to human health.

Methodology. The genome was sequenced, and a hybrid assembly of short and long reads produced. Antimicrobial susceptibility was determined by the broth microdilution method. Virulence was assessed by measuring *in vitro* biofilm formation compared to *Pseudomonas aeruginosa* and *in vivo* lethality in *Galleria mellonella* larvae.

Results. The isolate was confirmed to be a strain of *C. pauculus*, with a 6.8 Mb genome consisting of 6468 coding sequences and an overall G+C content of 63.9 mol%. The genome was found to contain 12 antibiotic resistance genes, 8 virulence factor genes and 33 metal resistance genes. The isolate can be categorized as resistant to meropenem, amoxicillin, amikacin, gentamicin and colistin, but susceptible to cefotaxime, cefepime, imipenem and ciprofloxacin. Clear biofilm formation was seen in all conditions over 72 h and exceeded that of *P. aeruginosa* when measured at 37 °C in R2A broth. Lethality in *G. mellonella* larvae over 48 h was relatively low.

Conclusion. The appearance of a multidrug-resistant strain of *C. pauculus* in a known pathogen reservoir within a clinical setting should be considered concerning. Further work should be completed to compare biofilm formation and *in vivo* virulence between clinical and environmental strains, to determine how easily environmental strains may establish human infection. Infection control teams and clinicians should be aware of the emerging nature of this pathogen and further work is needed to minimize the impact of contaminated hospital plumbing systems on patient outcomes.

Received 03 November 2021; Accepted 29 December 2021; Published 03 February 2022

Author affiliations: ¹School of Engineering, Computing and Mathematics, Faculty of Science and Engineering, University of Plymouth, Plymouth PL4 8AA, UK; ²School of Biomedical Sciences, Faculty of Health, University of Plymouth, Plymouth PL4 8AA, UK; ³Peninsula Dental School, Faculty of Health, University of Plymouth, Plymouth PL4 8AA, UK.

*Correspondence: James Butler, james.butler@plymouth.ac.uk

Keywords: Sink trap; *Cupriavidus pauculus*; MF1; whole genome sequencing; *Galleria mellonella*; biofilm.

Abbreviations: ANI, average nucleotide identity; ARG, antibiotic resistance gene; bp, base pair; CBA, columbia blood agar; CV, crystal violet; EPS, extracellular polymeric substances; EUCAST, European Committee on Antimicrobial Susceptibility Testing; LD₅₀, median lethal dose; MIC, minimum inhibitory concentration; MLST, multi-locus sequence typing; MRG, metal resistance gene; ONT, Oxford Nanopore Technologies; PBS, phosphate-buffered saline; R2A, Reasoner's 2A; RT, room temperature; TSB, tryptic soy broth; VF, virulence factor; WGS, whole genome sequencing.

This project has been deposited at DDBJ/ENA/GenBank under the accession JAIQWY000000000 within BioProject PRJNA762702. The version described in this paper is version JAIQWY010000000. The contig sequences can be found under accession numbers JAIQWY010000001–JAIQWY010000005. Raw sequence data have been deposited under accession numbers SRX13365403 and SRX13365404.

Four supplementary tables are available with the online version of this article.

001501 © 2022 The Authors



INTRODUCTION

The genus *Cupriavidus* is a member of the family *Burkholderiaceae* and consists of Gram-negative peritrichously flagellated bacilli. *Cupriavidus* species are primarily environmental organisms found in soil [1–3] but have also been isolated from a wastewater treatment plant [4], a groundwater remediation system [5], a hydrotherapy pool [6], bottled mineral water [7], nebulization solutions [8], and even the International Space Station [9, 10]. Many historical isolates were first identified as ‘CDC Group IVC-2’, which was later classified as a *Ralstonia* species [11], then as *Wautersia* [12], before finally being transferred to the genus *Cupriavidus* [13]. Certain *Cupriavidus* species such as *Cupriavidus pauculus* are emerging as opportunistic pathogens of interest [14] with multiple case reports of infection in immunocompromised [15–19] and immunocompetent [20–23] people as well as in those with cystic fibrosis [24–26], with evidence of nosocomial transmission [27–29]. A comprehensive summary of bacteraemia caused by *Cupriavidus* species reported that *C. pauculus* accounted for the vast majority (87%) of 23 reported cases across different countries [28].

The susceptibility of epidemiologically unrelated *Cupriavidus* spp. clinical strains to 20 antibiotics has been characterized [30], revealing widespread resistance to amoxicillin, amoxicillin-clavulanate, temocillin and aztreonam, relatively low (23%) susceptibility to ceftazidime, and higher susceptibility to ceftriaxone and cefotaxime (74 and 82%, respectively). Mixed susceptibility to colistin was reported, which is significant as the *Cupriavidus* genus was initially described as being colistin-susceptible [31] but *C. metallidurans* was later found to exhibit high expression of aminoarabinose transferase (ArnT) [32], a colistin resistance determinant, and *C. gilardii* may have been the origin of the *mcr-5* gene [30, 33].

Sink traps (also known as U-bends or P-traps) are sections of wastewater plumbing systems (WPS), both domestic and commercial, designed to trap water and prevent unwanted flow of sewer gases into the sink and surrounding environment. Sinks are present in virtually all hospital wards and patient rooms to encourage best practice regarding hand hygiene [34, 35], however these sink traps become heavily colonized with pathogenic bacteria and are an important reservoir of pathogens causing nosocomial infections [36–40]. The formation of biofilms, which are refractory to disinfection and facilitate long-term persistence of bacteria, poses an additional concern [41]. As well as acting as a reservoir, sinks are points of direct pathogen dispersal when they are used as bacteria escape by droplets and aerosols [42–45], and bacteria can also be carried on airflows within plumbing systems even between different floors of a building [46]. Furthermore, due to the realistic nature of sinks being used as an occasional point of disposal of patient material, pharmaceuticals and/or disinfectants, their resident bacteria are exposed to a wide variety of chemical substances. One study of an intensive care unit found that only 4% of total sink-related behaviours could be categorized as hand-washing, and a total of 56 sink-related activities could be identified that introduced a variety of nutrients potentially promoting microbial growth in the sinks [47]. There is evidence to suggest that hospital effluent wastewater selects for antibiotic resistance [48], suggesting that surfaces in the WPS may become enriched with resistant bacteria and so WPS could be further thought of as a reservoir of resistance genes. These resistance genes can reside in various genera of hardy environmental organisms and sinks may provide a platform for horizontal gene transfer [49]. *C. pauculus* has previously been linked with clinical WPS: in one case a *C. pauculus* bacteraemia was linked to a contaminated hospital water system [50] while in another case a ‘pseudo-outbreak’ at an outpatient clinic was linked to rinsing culture swabs in tap water [51], however isolation of *C. pauculus* from a hospital sink trap has not previously been reported.

In this article, we report the annotated draft genome sequence of *C. pauculus* strain MF1 isolated from a sink trap in a UK hospital. We also report the isolate’s genotypic and phenotypic antimicrobial susceptibility profile, determination of median lethal dose in *Galleria mellonella* larvae, and biofilm formation compared to *Pseudomonas aeruginosa*.

METHODS

Sample collection and bacterial recovery

Sink traps ($n=2$) were removed from sinks at a UK hospital in 2017 by hospital staff, sealed in individual sample bags, transferred to University of Plymouth laboratories, and stored at -20°C . In 2020, sink traps were thawed to room temperature (RT) for 1 h before sampling with a sterile swab moistened with Dulbecco A PBS (Thermo Scientific Oxoid, UK). Swabs were placed in tubes containing 3 ml PBS, then rotated and vortexed. Suspensions were serially diluted tenfold to 10^{-3} and all subsequent suspensions were spread plated in 100 μl aliquots on columbia blood agar (CBA) composed of columbia agar (Sigma-Aldrich, MO, USA) with 5% (v/v) defibrinated horse blood (Fisher Scientific, UK). All bacteriological media and PBS were sterilized by autoclaving at 121°C and 15 psi for 15 min. Agar plates were incubated at either 20°C or 37°C for up to 7 days.

Gram stain

To visualize cell morphology, indicate cell-wall composition and ultimately aid in initial identification of the organism, a Gram stain was conducted. A few colonies of the isolate were emulsified in a small volume of sterile deionized water spread across a glass slide, allowed to dry to give a smear, and fixed by briefly passing through a Bunsen flame. The slide was stained for 1 min by flooding with crystal violet (CV) solution (Pro-Lab Diagnostics, UK), rinsed with tap water, then flooded with the mordant

Gram's iodine (Pro-Lab Diagnostics, UK) for 1 min before rinsing briefly again with tap water. The slide was then held on a slant while a few drops of the decolourizing agent 95% ethanol were added and rinsed off with tap water. Finally, the counterstain safranin (Pro-Lab Diagnostics, UK) was added for 1 min and rinsed off with tap water. The slide was blotted dry and visualized with a brightfield microscope.

Strain identification by 16S rRNA gene sequencing

Identification was continued based on the 16S rRNA gene sequence. Genomic DNA was extracted from a pure culture using the DNeasy PowerSoil Kit (Qiagen, Germany), according to the manufacturer's instructions. This kit utilizes an initial bead-beating step to lyse cells, followed by binding to a silica membrane within a spin column in the presence of chaotropic salts, removal of contaminants, and finally elution of purified DNA from the membrane. The 16S rRNA gene was amplified by PCR in 50 µl reactions consisting of 25 µl DreamTaq Green 2× PCR Master Mix (Thermo Fisher Scientific, UK), 5 µl DNA template, 2.5 µl of each primer (Eurofins Genomics, Luxembourg) at 0.5 µM (27 f: 5'-AGA GTT TGA TCA TGG CTC A-3', 1492 r: 5'-TAC GGT TAC CTT GTT ACG ACT T-3'), and 15 µl Ambion nuclease-free water (Invitrogen, MA, USA). A positive control of a known bacterial DNA sample and a negative control of Ambion nuclease-free water were included in the place of the DNA template. The thermocycler programme was as follows: 5 min at 94 °C followed by 30 cycles of 30 s at 94 °C, 1 min at 52 °C, and 1.5 min at 72 °C, followed by 10 min at 72 °C. The presence and size of PCR products were confirmed by gel electrophoresis, running 5 µl of each PCR reaction mixture on a 1.5% agarose gel with SYBR Safe DNA Gel Stain (Invitrogen, MA, USA). PCR products were prepared for sequencing using the ExoSAP-IT PCR Product Cleanup Reagent (Thermo Fisher Scientific, UK) according to the manufacturer's instructions. Amplicon sequencing was completed by Biosearch Technologies (LGC Genomics, Germany), and sequences were analysed using Geneious Prime 2021.1.1 for macOS (<https://geneious.com>). The ends of the forward and reverse sequences were trimmed based on quality (error probability limit 0.01) and the sequences aligned using the MUSCLE algorithm (default settings) within Geneious Prime. The resulting consensus sequence was then used for a search using the Basic Local Alignment Search Tool (BLAST; <https://blast.ncbi.nlm.nih.gov/Blast.cgi>).

Antimicrobial susceptibility testing

The guidelines provided by the European Committee on Antimicrobial Susceptibility Testing (EUCAST; <https://www.eucast.org>) were followed for completion of MIC assays by the broth microdilution method. All MIC assays were conducted in cation-adjusted Mueller–Hinton II broth (Sigma-Aldrich, MO, USA) in 96-well round-bottom polypropylene microplates (Corning, NY, USA) and results determined visually following 18±2 h static incubation at 37 °C. Antibiotics were purchased from Melford Laboratories (amikacin, amoxicillin, amoxicillin-clavulanate, cefotaxime, colistin), Sigma-Aldrich (ceftazidime, ciprofloxacin, doxycycline, erythromycin, meropenem), Carbosynth (imipenem, relebactam), Fisher Scientific (cefepime) and Lonza (gentamicin). Antibiotic stocks were prepared in sterile deionized water, except erythromycin, which was prepared in dimethyl sulfoxide, and stored at -20 °C until use. A 16:1 ratio of amoxicillin to relebactam was used, as previously reported [52]. Classification of the isolate's susceptibility to antibiotics was determined based on EUCAST breakpoints [53] using entries for *Pseudomonas* spp. as published previously [28, 30, 54], as well as PK-PD breakpoints which are not species-specific.

Draft genome sequence determination

Illumina next-generation sequencing was provided by MicrobesNG (<https://microbesng.com>). The isolate was submitted in the form of cells harvested from an agar plate after streaking a single colony, suspended in a tube with cryopreservative (Microbank, Pro-Lab Diagnostics, UK). DNA extraction was completed by MicrobesNG using lysozyme, RNase A, proteinase K, and sodium dodecyl sulphate, and genomic DNA was purified using SPRI beads. Details of the DNA extraction, quantification and sequencing library preparation are available at <https://microbesng.com/microbesng-faq/>. Finally, reads were adapter-trimmed using Trimmomatic 0.30 with a sliding window quality cut-off of Q15.

Oxford Nanopore Technologies (ONT) MinION sequencing was employed to produce long reads to assist in genome assembly. Genomic DNA was isolated using the DNeasy Blood and Tissue Kit (Qiagen, Germany) with a modification made for Gram-negative bacteria according to the manufacturer's instructions; brief suspension in a lysis buffer (kit buffer ATL). Precautions were taken throughout to reduce shearing of high molecular weight DNA, including flicking tubes to mix rather than vortexing, use of wide bore pipette tips, avoidance of freeze-thaw cycles, and keeping tubes on ice. Extraction yield was quantified using the Qubit dsDNA HS Assay Kit (Invitrogen, MA, USA) and fluorometer (Thermo Fisher Scientific, UK), and purity was determined using a Nanodrop spectrophotometer (Thermo Fisher Scientific, UK). ONT sequencing was completed using the 1D Native barcoding genomic DNA protocol (SQK-LSK108) and EXP-NBD103 barcoding kit on an R9.4.1 flow cell (FLO-MIN106) according to the manufacturer's instructions. Basecalling was completed using Guppy version 5.0.11 with default settings using the Cloud Infrastructure for Big Data Microbial Bioinformatics (CLIMB) platform [55].

Hybrid genome assembly and annotation

A hybrid assembled and annotated genome consisting of Illumina short reads and ONT long reads was produced using the Comprehensive Genome Analysis tool provided by the PATRIC 3.6.10 Bioinformatics Resource Center [56]. Unicycler [57] was selected as the assembly strategy and annotation was completed using the RAST toolkit (RASTtk) [58].

In vivo *G. mellonella* infection to determine median lethal dose of *C. pauculus* MF1

Larvae of the greater wax moth *G. mellonella* were purchased from Livefood UK and stored in the dark at 4 °C for up to 7 days without food or water. Larvae were selected to be 21 ± 1 mm in length and healthy, as determined by a uniform cream colour, with no indications of melanization. Larval length can be correlated to weight according to a linear regression, such that a length of 21 ± 1 mm equates to a weight of 208.2 ± 22.2 mg, which is within the optimum range (180–260 mg) for determination of median lethal dose (LD_{50}) using this model [59]. To prepare the *C. pauculus* MF1 inoculum, the isolate was grown overnight in 10 ml LB broth (Fisher Scientific, UK) at 37 °C and collected by centrifugation at 2500 g for 10 min. The pellet was then resuspended in 10 ml PBS to wash the cells, before repeating the centrifugation once more and finally resuspending the pellet in 1 ml PBS. The resulting suspension was tenfold serially diluted and bacterial density was confirmed by viable count assay using the Miles and Misra method [60].

Ten larvae were injected per serial dilution of *C. pauculus* MF1 (10^2 , 10^3 , 10^4 , 10^5 , 10^6 and 10^7 c.f.u./10 µl) into the left penultimate pro-leg using a 50 µl Hamilton 750 syringe (Hamilton Company, NV, USA) with a removable 26S gauge needle. A placebo control of sterile PBS was used to account for the physical trauma of injection ($n=10$), along with a 'no manipulation' control to account for normal survival ($n=10$). Injected larvae were placed into sterile Petri dishes and incubated at 37 °C in the dark without food or water for 48 h. After 24 and 48 h, the percentage of live larvae in each treatment group was recorded. Larvae were recorded as dead when they met at least two of the following criteria: (i) obvious melanization, (ii) no response to touch, (iii) no correction when rolled on back [59]. For each time point, the LD_{50} was calculated by plotting c.f.u./larva against percentage live larvae and interpolating the curve for 50%. The mean average of three biological replicates is reported.

Biofilm formation compared to *P. aeruginosa* PAO1

The CV solubilization assay was used to measure the biomass of formed biofilm in microplates according to the methods published previously [61–63]. *P. aeruginosa* was selected as a comparator organism due to its wide acceptance as a versatile biofilm-forming organism [64–66], and PAO1 was selected due to its ubiquity in research as a well-characterized strain [67]. *C. pauculus* MF1 and *P. aeruginosa* PAO1 were each inoculated in tryptic soy broth (TSB; Sigma-Aldrich, MO, USA) and Reasoner's 2A (R2A) broth (prepared in-house identically to Oxoid CM0906 but omitting agar, see the Supplementary Material, available in the online version of this article, for composition) and incubated at 37 °C until log phase. Cultures were then adjusted to 1×10^6 c.f.u. ml⁻¹ and 200 µl/well added to flat-bottom 96-well plates (Thermo Scientific Biolite). There were 12 technical replicates per bacteria-media combination. Outer microplate wells were filled with 200 µl sterile water to exclude any edge effects and a negative control of broth alone was included. Plates were incubated statically at 37 °C or 20 °C for 24–72 h. To quantify biomass of biofilm formed, at each time point plates were washed with dH₂O to remove nonadherent cells, stained by addition of 200 µl/well 0.1% (*w/v*) CV solution, and incubated at RT for 20 min. CV was removed by pipette and then wells were washed again with dH₂O to remove unbound CV. Plates were dried at 60 °C for 30 min before cooling and solubilizing CV by addition of 200 µl/well 30% (*v/v*) acetic acid and incubating at RT on a shaking platform for 30 min. Absorbance was read at 575 nm using a SPECTROstar Omega (BMG Labtech, UK). The mean absorbance of negative controls was subtracted from that of each experimental well to account for background staining. At each time point, plates not being measured were also refreshed by removing liquid from wells and replacing with 200 µl/well appropriate fresh media.

Statistical analysis

All experiments consist of three independent biological replicates and at least three technical replicates per group, or more where indicated. Statistical analyses were conducted using GraphPad Prism v9.1.1 for macOS (GraphPad Software).

Data were checked for normal distribution using the Shapiro–Wilk test. Two-way ANOVA was conducted on biofilm data, with Šidák's multiple comparisons test. All statistical analyses used a 95% confidence limit, so that *P*-values equal to or greater than 0.05 were not considered statistically significant.

RESULTS

Initial isolation and identification of the strain

The isolate appeared as grey round easily emulsifiable colonies on CBA, and Gram-negative bacilli were apparent on staining. Sanger sequencing of the 16S rRNA gene PCR product and subsequent processing yielded a 1392 base pair (bp) consensus sequence. A BLAST search gave resolution to genus-level, but species-level identification could not be completed due to many

100% identity and 100% query cover matches to multiple *Cupriavidus* species (*C. pauculus*: MW960228, MW049071, MW049067, AY860236, AB109753; *C. taiwanensis*: EU915716) and multiple sequences identified only to genus level as *Cupriavidus* (GU566329, EU827490, EU596431).

Draft genome sequencing

Incorporation of both the Illumina short reads and ONT long reads in a hybrid assembly resulted in a 6.8 Mb draft genome of five contigs, with a 'good' quality determination by PATRIC and 0% contamination. The number of coding sequences (CDS) was 6468 – of which 4063 were given functional assignments. PATRIC indicated that the genome was 99.5% complete. For further details of the hybrid assembly and annotated genome, see Table S1. Summary statistics from the ONT MinION sequencing run can be found in Table S2 and characteristics of an assembly resulting from Illumina reads alone can be found in Table S3.

Multi-locus sequence typing (MLST) using the autoMLST web tool (<https://automlst.ziemertlab.com/>) gave the closest match as 98.2% average nucleotide identity (ANI) to *C. pauculus* KF709 (NBRC 110672) deposited as GCA_000974605 [3]. As the ANI cut-off for new prokaryotic species is given as 95% [68, 69], the isolate was concluded to be part of the existing species *C. pauculus*. See Table S4 for the top 10 MLST results. To confirm the MLST classification, the Genome-to-Genome Distance Calculator provided by the Leibniz Institute DSMZ was used to run a digital DNA–DNA hybridization (DDH) to mimic DDH *in silico* [70]. This tool uses three independent formulae, reporting three DDH values. The results were 73.8, 85.4 and 78.5% (mean 79.2%) against the closest match, *C. pauculus* KF709 as above. A DDH value of $\geq 70\%$ is recommended as the threshold for definition of members of the same species [71, 72], thus on this basis the isolate was confirmed to be part of the existing species *C. pauculus*. The strain was designated MF1.

The RASTtk within PATRIC was used to identify antibiotic resistance genes (ARGs), virulence factor genes (VFs) and metal resistance genes (MRGs). The full list of the genes of interest within these categories can be found in Table 1. In total, 12 ARGs, 8 VF genes and 33 MRGs were identified.

Susceptibility to antibiotics

The MICs of antibiotics against the *C. pauculus* MF1 isolate and the resulting susceptibility classifications can be found in Table 2. As breakpoint tables for this genus are not available, susceptibility classifications were based on breakpoints for *Pseudomonas* spp. as has been published previously [28, 30, 54], as well as PK-PD breakpoint values, which are not species-specific. The isolate showed susceptibility or intermediate susceptibility to the cephalosporins cefotaxime, ceftazidime and cefepime. For carbapenems, resistance to meropenem was clear however the isolate was susceptible to imipenem with little additional benefit conferred by the inclusion of the β -lactamase inhibitor relebactam. The isolate was resistant to amoxicillin but could be sensitized to some degree by the addition of the β -lactamase inhibitors relebactam or clavulanate, which reduced the MIC by over 32-fold and 8-fold, respectively. Low MICs were observed for both ciprofloxacin (a fluoroquinolone) and doxycycline (a tetracycline) while high MICs were found for both gentamicin and amikacin (aminoglycosides). The MIC of erythromycin was above that which could be considered clinically beneficial, however conclusive classification is not possible due to the lack of a published breakpoint. Finally, the isolate can be classified as resistant to colistin.

Median lethal dose in *G. mellonella* larvae

The 24 h LD₅₀ of *C. pauculus* MF1 in *G. mellonella* was determined to be 2.54×10^7 c.f.u./larva ($SD \pm 5.99 \times 10^6$) while the 48 h LD₅₀ was determined to be 2.64×10^6 c.f.u./larva ($SD \pm 1.82 \times 10^6$). Taking the calculated average weight of larvae used in this study as 208.2 mg, these LD₅₀ values equate to 1.22×10^5 c.f.u. mg⁻¹ (24 h) and 1.27×10^4 c.f.u. mg⁻¹ (48 h).

Biofilm formation compared to *P. aeruginosa* PAO1

Both *C. pauculus* MF1 and *P. aeruginosa* PAO1 formed detectable biofilms in minimal (R2A) and rich (TSB) broth at both physiological (37 °C) and environmental (20 °C) temperatures between 24–72 h (Fig. 1). In R2A broth at 37 °C, *C. pauculus* biofilm built gradually and significantly overtook that of *P. aeruginosa* after 48 and 72 h (Fig. 1a). In R2A broth at 20 °C, *P. aeruginosa* formed much more biofilm, almost reaching maximum signal after only 24 h, while *C. pauculus* biofilm steadily built across the whole 72 h, though not overtaking that of *P. aeruginosa* under the same conditions (Fig. 1b). Generally, biofilm levels reached in TSB were greater than in R2A broth (Fig. 1c, d). In TSB at 37 °C, the *C. pauculus* biofilm climbed from a low level at 24 h and reached a similar high level as *P. aeruginosa* after 72 h (Fig. 1c). In TSB at 20 °C, *P. aeruginosa* biofilm levels reached similar high levels after 48 h onwards but at 24 h showed clearly less biofilm than at 37 °C (Fig. 1d). The *C. pauculus* biofilm built across 24–72 h in TSB at 20 °C but did not reach the same level as *P. aeruginosa* by the endpoint (Fig. 1d).

DISCUSSION

C. pauculus is an emerging human pathogen, having previously been chiefly isolated from soil and aquatic environments. Here, we present its isolation from a hospital sink trap – a niche known to act as a reservoir of pathogens in clinical settings and

Table 1. Genes of interest found within the *C. pauculus* MF1 draft genome

Gene type	Gene name	Gene product/function
Antibiotic resistance genes	<i>AmpC</i>	β -lactamase
	<i>adeF, adeG</i>	RND multidrug efflux pump (AdeFGH)
	<i>arnT</i>	Aminoarabinose transferase enzyme conferring colistin resistance
	<i>bla_{OXA}</i>	Class D β -lactamase
	<i>ceoB</i>	Cytoplasmic membrane component of the CeoAB-OpcM efflux pump
	<i>EmrAB-TolC</i>	Multidrug efflux system
	<i>H-NS</i>	Regulates expression of multidrug exporter genes
	<i>MacA, MacB</i>	Form an antibiotic efflux complex with TolC active against macrolides
	<i>MdtABC-TolC</i>	Multidrug efflux system
	<i>MexAB-OprM</i>	Multidrug efflux system
Virulence factors	<i>argG</i>	Argininosuccinate synthase; arginine synthesis
	<i>aroC</i>	Chorismate synthase; aromatic amino acid synthesis
	<i>hfq</i>	RNA chaperone at the post-transcriptional level
	<i>leuB</i>	3-isopropylmalate dehydrogenase
	<i>ppsA</i>	Phosphoenolpyruvate synthase
	<i>recA</i>	DNA repair and maintenance
	<i>rpoE</i>	RNA polymerase sigma E; facilitates bacterial growth at high temperature and may facilitate persistence in macrophages
Metal resistance genes	<i>trpG</i>	Anthranilate synthase component
	<i>acr3</i>	Transmembrane arsenite transporter
	<i>arsH</i>	Methylarsenite oxidase; ferric reductase activity and protection from oxidative stress
	<i>cbtA</i>	Putative cobalt transporter subunit
	<i>cnrR, cnrY</i>	Nickel and cobalt resistance
	<i>cobS-T, cobN, cobW</i>	Cobalt chelatases involved in cobalamin synthesis, <i>cobW</i> is a putative zinc chaperone
	<i>copC-D, copZ</i>	Copper chaperone and sequestration, may also bind silver and cadmium
	<i>corA, corC</i>	Na ⁺ -dependent Mg ²⁺ transporter
	<i>cusA-C, cusR-S</i>	Copper and silver resistance
	<i>cutE</i>	Copper transport
	<i>czcA-D, czcI, czcN</i>	Cadmium, zinc, and cobalt efflux
	<i>merP, merR, merT</i>	Mercury resistance
	<i>modA-C</i>	Components of molybdate uptake operon
<i>nikR</i>	Transcriptional regulator of the nikABCDE operon	

Listed are genes of interest found in the *C. pauculus* MF1 isolate whole genome, as annotated and highlighted by the PATRIC Comprehensive Genome Analysis tool using both CARD and PATRIC databases.

contribute to nosocomial transmission [36–40]. This study reports the draft hybrid genome sequence of the isolate and investigates its antimicrobial susceptibility, virulence in an *in vivo* model, and biofilm formation in relevant conditions compared to a well-characterized biofilm-forming model organism.

The isolate could not be identified to species-level using 16S rRNA gene sequencing; WGS was therefore indicated for identification and to further investigate the organism. A hybrid assembly of ONT and Illumina reads allowed a more complete genome to be

Table 2. MICs of antibiotics against the *C. pauculus* MF1 isolate

Class	Antibiotic	MIC ($\mu\text{g ml}^{-1}$)	Classification (<i>Pseudomonas</i> spp.)	Classification (PK-PD)
Cephalosporins	Cefotaxime	1	N/A	Susceptible
	Ceftazidime	8	Intermediate*	Intermediate
	Cefepime	0.25	Intermediate*	Susceptible
Carbapenems	Meropenem	>512	Resistant	Resistant
	Imipenem	0.5	Intermediate*	Susceptible
	Imipenem-Relebactam (2:1)	0.25	Susceptible	Susceptible
Penicillins	Amoxicillin	>512	N/A	Resistant
	Amoxicillin-Relebactam (16:1)	16	N/A	N/A
	Amoxicillin-Clavulanate (5:1)	64	N/A	Resistant
Aminoglycosides	Gentamicin	512	IE	Resistant
	Amikacin	512	Resistant	Resistant
Fluoroquinolones	Ciprofloxacin	0.125	Intermediate*	Susceptible
Macrolides	Erythromycin	32	N/A	IE
Tetracyclines	Doxycycline	0.25	N/A	IE
Polymyxins	Colistin	16	Resistant	IE

MICs of antibiotics against the *C. pauculus* MF1 isolate and the resulting susceptibility/resistance classifications. Classifications according to the breakpoints for *Pseudomonas* spp. are given as well as classifications based on pharmacokinetic-pharmacodynamic data provided by EUCAST. 'N/A' indicates that a breakpoint is not available. 'IE' indicates that EUCAST report there is insufficient evidence to consider a particular agent as a therapy and give no breakpoint. Intermediate with an asterisk (*) indicates that an MIC breakpoint of $\leq 0.001 \mu\text{g ml}^{-1}$ is given as an 'arbitrary off scale' breakpoint by EUCAST. Where any set of results contained differences of more than one serial dilution step, the assays were repeated. Shown are the consensus (mode) results of three biological replicates; alternatively, the midpoint was used if the three results were within one serial dilution step of each other.

achieved, adding to only six other genomes available for this species – two of which are PacBio/Illumina hybrids. Of these hybrids, both (GCA_008693385 and GCA_003854935) were submitted as part of the same FDA-ARGOS [73] project (PRJNA231221) in 2019 and consist of two contigs with one and two plasmids, respectively. The number of CDS identified in this study (6468) was greater than in both available hybrid genomes (6286 and 5483) although there was a difference in annotation method (RASTtk in PATRIC for this study vs NCBI Prokaryotic Genome Annotation Pipeline for the FDA-ARGOS project). There was also a difference in total sequence length (6.8 Mb versus 7.1 Mb and 6.2 Mb). Finally, the genome reported in this study did not contain any identifiable plasmids.

Through WGS, several ARGs were identified (Table 1) – mostly components of efflux pumps (*adeG*, *EmrAB-TolC*, *MdtABC-TolC*, *MexAB-OprM*, *MacA/MacB*) as well as other sequences identified by RASTtk as β -lactamases (*AmpC*, *bla_{OXA}*). Phenotypic resistance to meropenem, amoxicillin, amikacin, gentamicin and colistin was evident. The β -lactamase inhibitors relebactam and clavulanate reduced the MIC of amoxicillin, suggesting a resistance mechanism involving a β -lactamase, however the resulting MICs were still not within a clinically relevant range. The isolate was clearly susceptible to cefotaxime, cefepime, imipenem and ciprofloxacin. The antibiotic susceptibility profile appeared to broadly match that for *C. pauculus* reported by Massip *et al.* [30], except in the cases of meropenem (>512 $\mu\text{g ml}^{-1}$ here versus 16–64 $\mu\text{g ml}^{-1}$ reported) and amikacin (512 $\mu\text{g ml}^{-1}$ here versus 8–128 $\mu\text{g ml}^{-1}$ reported). Through further analysis of the ARGs identified by RASTtk, it became apparent that certain genes, which only confer resistance following acquisition of point mutations, such as *gyrA* and *rpsJ* [74, 75], are automatically identified as genes of interest – even when they lack the required mutations. Following investigation, these genes were discounted from our reported genes of interest. This highlights the importance of manually checking the output of genome annotation pipelines to confirm the relevance of identified genes.

While none of the mobilized colistin resistance (*mcr*) genes were identified in the genome, the isolate's phenotypic resistance to colistin may be explained by the presence of an *arnT* homologue with 81% identity and 99% query cover against that reported in *C. metallidurans* CH34 (CP000353.2:1481129–1482725) [30]. Aminoarabinose transferase (ArnT) catalyses the attachment of the cationic sugar 4-amino-4-deoxy-L-arabinose to lipid A, resulting in charge modification of the bacterial outer membrane and polymyxin resistance [32]. Homologues of *arnT* have previously been described in *C. pauculus*, *C. basilensis*, *C. necator* and *C. taiwanensis* as the cause of colistin resistance [30].

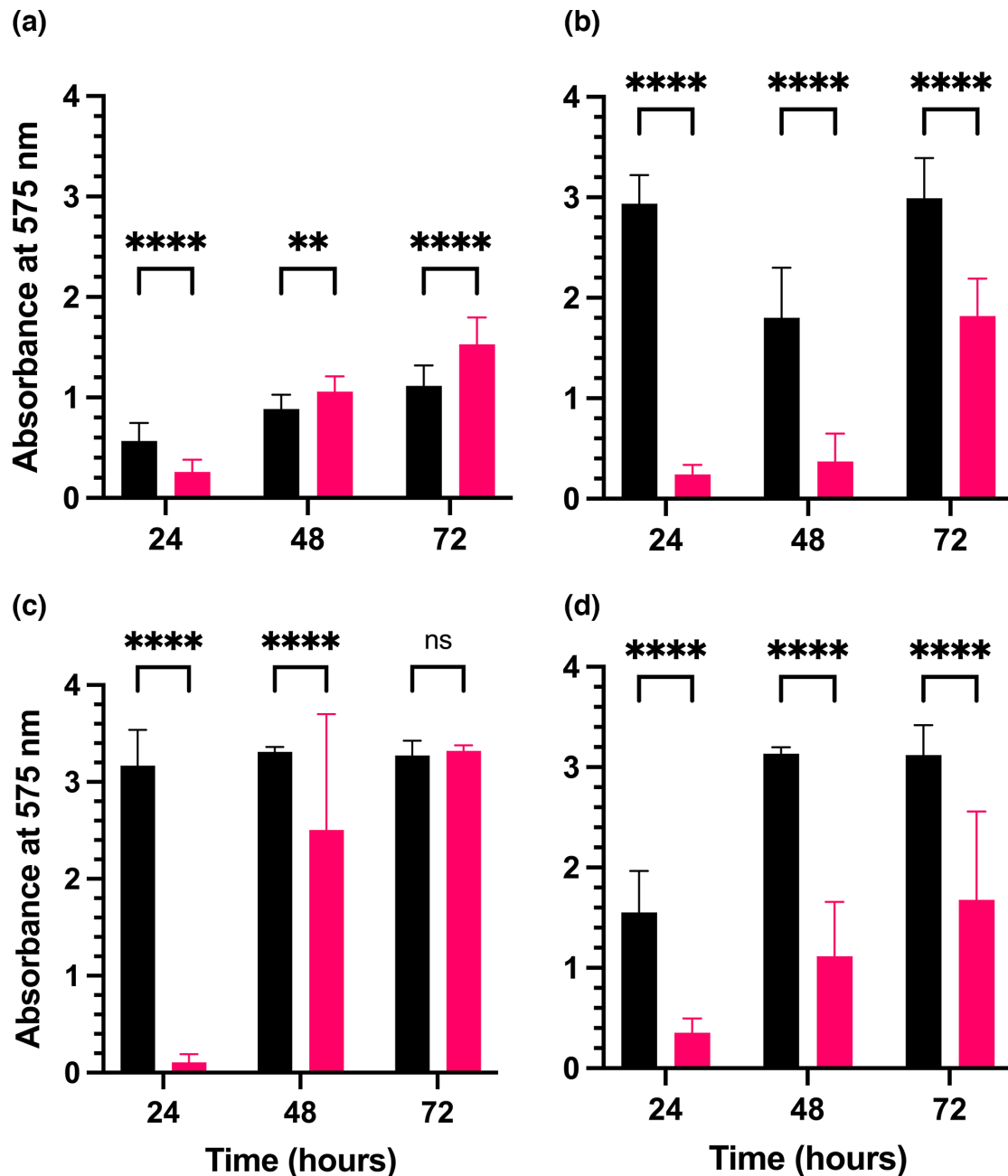


Fig. 1. Biofilm formation by *C. pauculus* MF1 (pink) and *P. aeruginosa* PA01 (black) in R2A broth at either 37 °C (a) or 20 °C (b) and in TSB at either 37 °C (c) or 20 °C (d) over 72 h as determined by the CV solubilization assay. Data presented as mean \pm SD. **** $P < 0.0001$, ** $P < 0.01$, ns: not significant; two-way ANOVA with Šidák's multiple comparisons test. Data presented are the average of $n=3$ biological replicates.

A large number ($n=33$) of MRGs were identified in the WGS. Together, the MRGs listed in Table 1 contribute to resistance to copper, silver, magnesium, cobalt, mercury, molybdenum, arsenic, zinc, cadmium, iron, antimony and nickel. Many species of *Cupriavidus* including *C. pauculus* [76, 77], *C. metallidurans* [78, 79], *C. gilardii* [80], *C. campinensis* [81] and *C. neocaledonicus* [82] are considered to be metal resistant. Many of these genes confer resistance to metals by means of intracellular metal sequestration to facilitate export, as in the case of *CopZ* with Cu^+ ions [83], Ag^+ and Cd^{2+} ions [84]. There is a link between metal and antibiotic resistance, in that ARGs can co-locate on mobile genetic elements carrying MRGs such as the integrative conjugative element ICEHs1, which contains 83 genes, including both ARGs (*tetH*, *ebrB*) and MRGs (*mco*, *czcD*, *acr3*) [85].

The RASTtk annotation also identified a number ($n=8$) of VFIs. One of these, *hfq*, is a known pleiotropic regulator of virulence genes with the protein Hfq acting as an RNA chaperone and modulating mRNA translation and stability [86]. Another, *recA*, is involved in

recombination, repair and maintenance of DNA, as well as induction of prophage [87]. *Salmonella typhimurium* mutants deficient of *recA* have been shown to be less virulent in BALB/c mice and more susceptible to macrophage killing by oxidative burst [88]. The alternative sigma factor *rpoE* is essential for some species such as *E. coli* [89], and is also induced by stressors, including heat and accumulation of misfolded proteins, to trigger extensive alterations in gene expression [90, 91]. RpoE, its product, has been shown to be important for intracellular survival of *Salmonella typhimurium* in macrophages [92]. The *G. mellonella* larvae model, a well-characterized model for virulence and lethality testing, was used to assess median lethal dose of the isolate. Non-lethal isolates (non-pathogenic *E. coli* DH5a) have previously been reported with an LD₅₀ of around 10⁷ c.f.u./larva [93], which is around the same range as the 24h LD₅₀ values reported in this study. As would be expected, the 48h LD₅₀ values were lower than those at 24h but remained relatively high. This suggests that *C. pauculus* MF1 exhibits low levels of virulence, at least in *G. mellonella* in the conditions described. There is some room for interpretation in all findings for *G. mellonella* as authors calculate and report results in myriad ways, e.g. c.f.u./larva versus c.f.u. mg⁻¹, and some authors do not report whether they controlled for larval size or weight. Larval weight shows a strong correlation to larval liquid volume [94], and it follows that injections into larger larvae will lead to greater inoculum dilution possibly leading to error in the reported LD₅₀. *G. mellonella* are particularly susceptible to killing by *P. aeruginosa*, with a reported LD₅₀ of only 10 c.f.u. [95]. Meanwhile the 24h LD₅₀ values of uropathogenic *E. coli* isolates have been determined to be around 1.6×10⁴ c.f.u./larva (ST69) and 1.2×10⁴ c.f.u./larva (ST127) [93]. The 48h LD₅₀ of two strains of *Burkholderia cepacia* have been reported as 30 c.f.u./larva or 1 c.f.u./larva while that of eight strains of *B. cenocepacia* ranged from 9×10² c.f.u./larva to 2×10⁵ c.f.u./larva [96]. It is noteworthy that despite them both belonging to *Burkholderiaceae*, *C. pauculus* MF1 and the strains of *B. cepacia* and *B. cenocepacia* have significantly different LD₅₀ values in *G. mellonella*, which may be explained by differences in virulence factors and resulting pathogenicity. The virulence of *Burkholderia* spp. is the result of potent biofilm formation, intrinsic antibiotic resistance, metabolic and genetic plasticity, multiple secretion systems, and the ability to persist intracellularly within macrophages and other eukaryotic cells [97]. Further evaluation of the *in vivo* virulence of this and other *C. pauculus* strains in another model would provide a valuable insight.

Biofilm formation was determined in 96-well plates according to a widely used protocol involving staining and solubilization of CV, which binds to all microbial biomass within each well. The CV solubilization assay therefore quantifies biofilm biomass, but cannot enumerate bacterial cells or indicate their viability. R2A agar was initially developed for enumeration and subculture of bacteria from potable water [98], and so we propose that R2A broth can serve as an appropriate medium for measuring biofilm formation in conditions most relevant to the sink trap and WPS as has been completed previously [99, 100]. However, as mentioned in the Introduction, it is important to bear in mind that disposal of liquids and solids via sink traps will result in variable nutrient conditions and so there is no perfect laboratory model to mimic real-life sink traps or WPS. At both physiological and environmental temperatures in minimal and rich conditions, *C. pauculus* biofilm was evident from 24h and increased consistently over the 72h experiments. When studied at 37°C in R2A broth, the *C. pauculus* MF1 biofilm even overtook that of *P. aeruginosa* PAO1 – a classic model organism for biofilm formation. There has been little previous work published on *C. pauculus* biofilm formation and, to our knowledge, this is the first report using the CV solubilization assay. A study investigating the efficacy of different disinfectants against biofilms containing *C. pauculus* recovered from the International Space Station's water system found that an 18h flush of either 6% H₂O₂ solution or a mixture of 3% H₂O₂ and 400 parts per billion colloidal silver effectively reduced the viability of 5 day preformed biofilms to <1 c.f.u. ml⁻¹ for up to 3 months. It is noteworthy that in the same study, the control biofilm became predominated with *C. pauculus* at an 8:2 ratio with *Burkholderia multivorans*, despite equal seeding of these and two other isolates [101]. The ability of *C. pauculus* to persist in oligotrophic conditions, whilst producing EPS, which may shield it from disinfectants and detergents, reinforces concerns that it may successfully persist in hospital WPS. The ability of *C. pauculus* MF1 to form a biofilm shown in this study, overtaking that of *P. aeruginosa* PAO1 at 37°C, is concerning and could suggest an ability to establish chronic, recalcitrant infections in humans like *P. aeruginosa* does, particularly in cystic fibrosis patients [65, 102]. It would be valuable to further investigate *C. pauculus* virulence under physiological conditions *in vivo*, perhaps with comparisons between clinical and environmental strains to determine whether potent biofilm formation is a feature of both, or is limited to environmental persistence.

This study cannot identify the source of the *C. pauculus* isolate recovered, thus it is impossible to determine whether the isolate described here originated from a human carrier, or was brought in via another route (e.g. the water supply). Nevertheless, it is concerning that an organism previously described as being of environmental origin, which has recently been implicated in several serious cases of human infection, has been isolated in a known pathogen reservoir in a clinical setting. It is further concerning that the isolate is multidrug resistant, including to the antibiotic of last-resort, colistin, to which it has previously been reported to be susceptible and which has been used therapeutically [17]. While there is clear evidence of *C. pauculus* causing morbidity and mortality, which makes its colonization of a sink trap a problem in its own right, it also poses a potential threat due to the carriage and possible horizontal transfer of ARGs. Further investigation and evidence is needed to be able to determine whether *C. pauculus* has more widely crossed into pathogen reservoirs such as sink traps, or whether the isolate described here was an incidental finding.

Funding information

This work received no specific grant from any funding agency. The authors acknowledge internal funding from the School of Engineering, Computing and Mathematics and the School of Biomedical Sciences at the University of Plymouth, which indirectly supported this work.

Acknowledgements

The authors are grateful to Dr Jonathan Cox (Aston University) and Mr Matthew Emery (University of Plymouth) for the provision of reagents for antimicrobial susceptibility testing, and to Dr Jo Fothergill (University of Liverpool) for the provision of *P. aeruginosa* PAO1. The authors thank Matthew Koch (University of Plymouth) and Poppy Hesketh Best (University of Plymouth) for advice on conducting Oxford Nanopore MinION sequencing.

Author contributions

Conceptualization: J.B., S.D.K. and M.U. Data curation: J.B. Formal analysis: J.B. Funding acquisition: M.U. and A.B. Investigation: J.B., S.D.K. and K.J.M. Methodology: J.B., S.D.K. and M.U. Project administration: M.U. Resources: J.B., S.D.K. and M.U. Supervision: M.U. Validation: J.B. and K.J.M. Visualization: J.B. Writing – original draft: J.B. Writing – review and editing: J.B., S.D.K., K.J.M., A.B. and M.U.

Conflicts of interest

The authors declare that there are no conflicts of interest.

References

- Feng T, Kim KH, Oh J, Jeon CO. *Cupriavidus lacunae* sp. nov., isolated from pond-side soil. *Antonie van Leeuwenhoek* 2019;112:543–551.
- Makkar NS, Casida LE. *Cupriavidus necator* gen. nov., sp. nov.; a nonobligate bacterial predator of bacteria in sNonobligate Bacterial Predator of Bacteria in Soil. *Int J Syst Bacteriol* 1987;37:323–326.
- Watanabe T, Yamazoe A, Hosoyama A, Fujihara H, Suenaga H, et al. Draft genome sequence of *Cupriavidus pauculus* strain KF709, a biphenyl-utilizing bacterium isolated from biphenyl-contaminated soil. *Genome Announc* 2015;3:e00222–15.
- Schmidt T, Schlegel HG. Nickel and cobalt resistance of various bacteria isolated from soil and highly polluted domestic and industrial wastes. *FEMS Microbiol Ecol* 1989;62:315–328.
- Wyndham RC, Nakatsu C, Peel M, Cashore A, Ng J, et al. Distribution of the catabolic transposon Tn5271 in a groundwater bioremediation system. *Appl Environ Microbiol* 1994;60:86–93.
- Aspinall ST, Graham R. Two sources of contamination of a hydrotherapy pool by environmental organisms. *J Hosp Infect* 1989;14:285–292.
- Manaia CM, Nunes OC, Morais PV, da Costa MS. Heterotrophic plate counts and the isolation of bacteria from mineral waters on selective and enrichment media. *J Appl Bacteriol* 1990;69:871–876.
- Oie S, Makieda D, Ishida S, Okano Y, Kamiya A. Microbial contamination of nebulization solution and its measures. *Biol Pharm Bull* 2006;29:503–507.
- Mora M, Perras A, Alekhova TA, Wink L, Krause R, et al. Resilient microorganisms in dust samples of the International Space Station-survival of the adaptation specialists. *Microbiome* 2016;4:65.
- Mijnendonckx K, Provoost A, Ott CM, Venkateswaran K, Mahillon J, et al. Characterization of the survival ability of *Cupriavidus metallidurans* and *Ralstonia pickettii* from space-related environments. *Microb Ecol* 2013;65:347–360.
- Moissenet D, Goujon CP, Garbarg-Chenon A, Vu-Thien H. CDC group IV c-2: a new *Ralstonia* species close to *Ralstonia eutropha*. *J Clin Microbiol* 1999;37:1777–1781.
- Vanechoutte M, Kämpfer P, De Baere T, Falsen E, Verschraegen G. *Wautersia* gen. nov., a novel genus accommodating the phylogenetic lineage including *Ralstonia eutropha* and related species, and proposal of *Ralstonia* [Pseudomonas] *syzygii* (Roberts et al. 1990) comb. nov. *Int J Syst Evol Microbiol* 2004;54:317–327.
- Vandamme P, Coenye T. Taxonomy of the genus *Cupriavidus*: a tale of lost and found. *Int J Syst Evol Microbiol* 2004;54:2285–2289.
- Langevin S, Vincelette J, Bekal S, Gaudreau C. First case of invasive human infection caused by *Cupriavidus metallidurans*. *J Clin Microbiol* 2011;49:744–745.
- Karafin M, Romagnoli M, Fink DL, Howard T, Rau R, et al. Fatal infection caused by *Cupriavidus gilardii* in a child with aplastic anemia. *J Clin Microbiol* 2010;48:1005–1007.
- Tena D, Losa C, Medina MJ, Sáez-Nieto JA. Muscular abscess caused by *Cupriavidus gilardii* in a renal transplant recipient. *Diagn Microbiol Infect Dis* 2014;79:108–110.
- Almasy E, Szederjesi J, Rad P, Georgescu A. A fatal case of community acquired *Cupriavidus Pauculus* pneumonia. *J Crit Care Med (Targu Mures)* 2016;2:201–204.
- Bianco G, Boattini M, Audisio E, Cavallo R, Costa C. Septic shock due to meropenem- and colistin-resistant *Cupriavidus pauculus*. *J Hosp Infect* 2018;99:364–365.
- Kweon OJ, Lim YK, Kim HR, Kim T-H, Ha S-M, et al. Isolation of a novel species in the genus *Cupriavidus* from a patient with sepsis using whole genome sequencing. *PLoS One* 2020;15:e0232850.
- Duggal S, Gur R, Nayar R, Rongpharpi SR, Jain D, et al. *Cupriavidus pauculus* (*Ralstonia paucula*) concomitant meningitis and septicemia in a neonate: first case report from India. *Indian J Med Microbiol* 2013;31:405–409.
- Kobayashi T, Nakamura I, Fujita H, Tsukimori A, Sato A, et al. First case report of infection due to *Cupriavidus gilardii* in a patient without immunodeficiency: a case report. *BMC Infect Dis* 2016;16:493.
- Yahya R, Alyousef W, Omara A, Alamoudi S, Alshami A, et al. First case of pneumonia caused by *Cupriavidus pauculus* in an infant in the Gulf Cooperation Council. *J Infect Dev Ctries* 2017;11:196–198.
- Shenai MB, Falconer R, Rogers S. A cupriavidus pauculus infection in a patient with a deep brain stimulation implant. *Cureus* 2019;11:e6104.
- Coenye T, Vandamme P, LiPuma JJ. Infection by *Ralstonia* species in cystic fibrosis patients: identification of *R. pickettii* and *R. mannitolilytica* by polymerase chain reaction. *Emerg Infect Dis* 2002;8:692–696.
- Kalka-MollWM, LiPumaJJ, AccursoFJ, PlumG, vanKoningsbruggenS, et al. Airway infection with a novel *Cupriavidus* species in persons with cystic fibrosis. *J Clin Microbiol* 2009;47:3026–3028.
- Lipuma JJ. The changing microbial epidemiology in cystic fibrosis. *Clin Microbiol Rev* 2010;23:299–323.
- Stovall SH, Wisdom C, McKamie W, Ware W, Dedman H, et al. Nosocomial transmission of *Cupriavidus pauculus* during extracorporeal membrane oxygenation. *ASAIO J* 2010;56:486–487.
- D’Inzeo T, Santangelo R, Fiori B, De Angelis G, Conte V, et al. Catheter-related bacteremia by *Cupriavidus metallidurans*. *Diagn Microbiol Infect Dis* 2015;81:9–12.
- Huda SA, Yadava S, Kahlown S, Jilani MH, Sharma B. A rare case of ventilator-associated pneumonia caused by *Cupriavidus pauculus*. *Cureus* 2020;12:e8573.
- Massip C, Coullaud-Gamel M, Gaudru C, Amoureux L, Doleans-Jordheim A, et al. In vitro activity of 20 antibiotics against *Cupriavidus* clinical strains. *J Antimicrob Chemother* 2020;75:1654–1658.
- Vanechoutte M, Kämpfer P, De Baere T, Falsen E, Verschraegen G. *Wautersia* gen. nov., a novel genus accommodating the phylogenetic lineage including *Ralstonia eutropha* and related species, and proposal of *Ralstonia* [Pseudomonas] *syzygii* (Roberts et al. 1990) comb. nov. *Int J Syst Evol Microbiol* 2004;54:317–327.
- Petrou VI, Herrera CM, Schultz KM, Clarke OB, Vendome J, et al. Structures of aminoarabinose transferase ArnT suggest a molecular basis for lipid A glycosylation. *Science* 2016;351:608–612.
- Zhang H, Zong Z, Lei S, Srinivas S, Sun J, et al. A genomic, evolutionary, and mechanistic study of MCR-5 action suggests

- functional unification across the MCR family of colistin resistance. *Adv Sci (Weinh)* 2019;6:1900034.
34. Rashid M. A decade of adult intensive care unit design: a study of the physical design features of the best-practice examples. *Crit Care Nurs Q* 2006;29:282–311.
 35. Hopman J, Tostmann A, Wertheim H, Bos M, Kolwijck E, et al. Reduced rate of intensive care unit acquired gram-negative bacilli after removal of sinks and introduction of “water-free” patient care. *Antimicrob Resist Infect Control* 2017;6:59.
 36. Lowe C, Willey B, O’Shaughnessy A, Lee W, Lum M, et al. Outbreak of extended-spectrum β -lactamase-producing *Klebsiella oxytoca* infections associated with contaminated handwashing sinks(1). *Emerg Infect Dis* 2012;18:1242–1247.
 37. Regev-Yochay G, Smollan G, Tal I, Pinas Zade N, Haviv Y, et al. Sink traps as the source of transmission of OXA-48-producing *Serratia marcescens* in an intensive care unit. *Infect Control Hosp Epidemiol* 2018;39:1307–1315.
 38. Volling C, Ahangari N, Bartoszko JJ, Coleman BL, Garcia-Jeldes F, et al. Are sink drainage systems a reservoir for hospital-acquired gammaproteobacteria colonization and infection? a systematic review. *Open Forum Infect Dis* 2021;8:faa590.
 39. Franco LC, Tanner W, Ganim C, Davy T, Edwards J, et al. A microbiological survey of handwashing sinks in the hospital built environment reveals differences in patient room and healthcare personnel sinks. *Sci Rep* 2020;10:8234.
 40. De Geyter D, Vanstokstraeten R, Crombé F, Tommassen J, Wybo I, et al. Sink drains as reservoirs of VIM-2 metallo- β -lactamase-producing *Pseudomonas aeruginosa* in a Belgian intensive care unit: relation to patients investigated by whole-genome sequencing. *J Hosp Infect* 2021;115:75–82.
 41. Ledwoch K, Robertson A, Lauran J, Norville P, Maillard JY. It’s a trap! The development of a versatile drain biofilm model and its susceptibility to disinfection. *J Hosp Infect* 2020;106:757–764.
 42. Hota S, Hirji Z, Stockton K, Lemieux C, Dedier H, et al. Outbreak of multidrug-resistant *Pseudomonas aeruginosa* colonization and infection secondary to imperfect intensive care unit room design. *Infect Control Hosp Epidemiol* 2009;30:25–33.
 43. Kotay S, Chai W, Guilford W, Barry K, Mathers AJ. Spread from the Sink to the Patient: *In Situ* Study Using Green Fluorescent Protein (GFP)-expressing *Escherichia coli* to model bacterial dispersion from hand-washing sink-trap reservoirs. *Appl Environ Microbiol* 2017;83:e03327–03316.
 44. Hajar Z, Mana TSC, Cadnum JL, Donskey CJ. Dispersal of gram-negative bacilli from contaminated sink drains to cover gowns and hands during hand washing. *Infect Control Hosp Epidemiol* 2019;40:460–462.
 45. Yui S, Muzslay M, Karia K, Shuttleworth B, Ali S, et al. Evaluation of droplet production by a new design of clinical handwash basin for the healthcare environment. *J Hosp Infect* 2019;103:e110–e114.
 46. Gormley M, Aspray TJ, Kelly DA, Rodriguez-Gil C. Pathogen cross-transmission via building sanitary plumbing systems in a full scale pilot test-rig. *PLoS One* 2017;12:e0171556.
 47. Grabowski M, Lobo JM, Gunnell B, Enfield K, Carpenter R, et al. Characterizations of handwashing sink activities in a single hospital medical intensive care unit. *J Hosp Infect* 2018;100:e115–e122.
 48. Kraupner N, Hutinel M, Schumacher K, Gray DA, Genheden M, et al. Evidence for selection of multi-resistant *E. coli* by hospital effluent. *Environ Int* 2021;150:106436.
 49. Muzslay M, Moore G, Alhussaini N, Wilson APR. ESBL-producing Gram-negative organisms in the healthcare environment as a source of genetic material for resistance in human infections. *J Hosp Infect* 2017;95:59–64.
 50. Inkster T, Peters C, Wafer T, Holloway D, Makin T. Investigation and control of an outbreak due to a contaminated hospital water system, identified following a rare case of *Cupriavidus pauculus* bacteraemia. *J Hosp Infect* 2021;111:53–64.
 51. Balada-Llasat J-M, Elkins C, Swyers L, Bannerman T, Pancholi P. Pseudo-outbreak of *Cupriavidus pauculus* infection at an outpatient clinic related to rinsing culturette swabs in tap water. *J Clin Microbiol* 2010;48:2645–2647.
 52. Lopeman RC, Harrison J, Rathbone DL, Desai M, Lambert PA, et al. Effect of amoxicillin in combination with imipenem-relebactam against *Mycobacterium abscessus*. *Sci Rep* 2020;10:928.
 53. EUCAST. *Breakpoint tables for interpretation of MICs and zone diameters*. 2021.
 54. Langevin S, Vincelette J, Bekal S, Gaudreau C. First case of invasive human infection caused by *Cupriavidus metallidurans*. *J Clin Microbiol* 2011;49:744–745.
 55. Connor TR, Loman NJ, Thompson S, Smith A, Southgate J, et al. CLIMB (the Cloud Infrastructure for Microbial Bioinformatics): an online resource for the medical microbiology community. *Microb Genom* 2016;2:e000086.
 56. Davis JJ, Wattam AR, Aziz RK, Brettin T, Butler R, et al. The PATRIC Bioinformatics Resource Center: expanding data and analysis capabilities. *Nucleic Acids Res* 2020;48:D606–D612.
 57. Wick RR, Judd LM, Gorrie CL, Holt KE. Unicycler: Resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 2017;13:e1005595.
 58. Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, et al. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 2015;5:8365.
 59. Hesketh-Best PJ, Mouritzen MV, Shandley-Edwards K, Billington RA, Upton M. *Galleria mellonella* larvae exhibit a weight-dependent lethal median dose when infected with methicillin-resistant *Staphylococcus aureus*. *Pathog Dis* 2021;79:ftab003.
 60. Miles AA, Misra SS, Irwin JO. The estimation of the bactericidal power of the blood. *J Hyg (Lond)* 1938;38:732–749.
 61. Wand ME, Bock LJ, Turton JF, Nugent PG, Sutton JM. *Acinetobacter baumannii* virulence is enhanced in *Galleria mellonella* following biofilm adaptation. *J Med Microbiol* 2012;61:470–477.
 62. O’Toole GA. Microtiter dish biofilm formation assay. *J Vis Exp* 2011;2011:47.
 63. Shukla SK, Rao TS. An improved crystal violet assay for biofilm-quantification in 96-well microtitre plate. *bioRxiv*; 100214 2017. . DOI: 10.1101/100214DOI: 10.1101/100214 [Preprint]
 64. Moradali MF, Ghods S, Rehm BHA. *Pseudomonas aeruginosa* Lifestyle: A Paradigm for Adaptation, Survival, and Persistence. *Front Cell Infect Microbiol* 2017;7:39.
 65. Høiby N, Ciofu O, Bjarnsholt T. *Pseudomonas aeruginosa* biofilms in cystic fibrosis. *Future Microbiol* 2010;5:1663–1674.
 66. Ciofu O, Tolker-Nielsen T. Tolerance and resistance of *Pseudomonas aeruginosa* biofilms to antimicrobial agents-how *P. aeruginosa* can escape antibiotics. *Front Microbiol* 2019;10:913.
 67. Klockgether J, Munder A, Neugebauer J, Davenport CF, Stanke F, et al. Genome diversity of *Pseudomonas aeruginosa* PAO1 laboratory strains. *J Bacteriol* 2010;192:1113–1121.
 68. Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, et al. DNA-DNA hybridization values and their relationship to whole-genome sequence similarities. *Int J Syst Evol Microbiol* 2007;57:81–91.
 69. Richter M, Rosselló-Móra R. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci U S A* 2009;106:19126–19131.
 70. Meier-Kolthoff JP, Auch AF, Klenk H-P, Göker M. Genome sequence-based species delimitation with confidence intervals and improved distance functions. *BMC Bioinformatics* 2013;14:60.
 71. Brenner DJ. Deoxyribonucleic acid reassociation in the taxonomy of enteric bacteria. *Int J Syst Bacteriol* 1973;23:298–307.
 72. Johnson JL. Use of nucleic-acid homologies in the taxonomy of anaerobic bacteria. *Int J Syst Bacteriol* 1973;23:308–315.
 73. Sichtig H, Minogue T, Yan Y, Stefan C, Hall A, et al. FDA-ARGOS is a database with public quality-controlled reference genomes for diagnostic use and regulatory science. *Nat Commun* 2019;10:3313.

74. Drlica K, Zhao X. DNA gyrase, topoisomerase IV, and the 4-quinolones. *Microbiol Mol Biol Rev* 1997;61:377–392.
75. Kubanov A, Vorobyev D, Chestkov A, Leinsoo A, Shaskolskiy B, et al. Molecular epidemiology of drug-resistant *Neisseria gonorrhoeae* in Russia (Current Status, 2015). *BMC Infect Dis* 2016;16:389.
76. Ramirez M, Obrzydowski J, Ayers M, Virparia S, Wang M, et al. Pyruvic oxime nitrification and copper and nickel resistance by a *Cupriavidus pauculus*, an active heterotrophic nitrifier-denitrifier. *ScientificWorldJournal* 2014;2014:901702.
77. Putonti C, Polley N, Castignetti D. Draft genome sequence of an active heterotrophic nitrifier-denitrifier, *Cupriavidus pauculus* UM1. *Genome Announc* 2018;6:e00028-18.
78. von Rozycki T, Nies DH. *Cupriavidus metallidurans*: evolution of a metal-resistant bacterium. *Antonie van Leeuwenhoek* 2009;96:115–139.
79. Mazhar SH, Herzberg M, Ben Fekih I, Zhang C, Bello SK, et al. Comparative insights into the complete genome sequence of highly metal resistant *Cupriavidus metallidurans* strain BS1 isolated from a gold-copper mine. *Front Microbiol* 2020;11:47.
80. Huang N, Mao J, Hu M, Wang X, Huo M. Responses to copper stress in the metal-resistant bacterium *Cupriavidus gilardii* CR3: a whole-transcriptome analysis. *J Basic Microbiol* 2019;59:446–457.
81. Abbaszade G, Szabó A, Vajna B, Farkas R, Szabó C, et al. Whole genome sequence analysis of *Cupriavidus campinensis* S14E4C, a heavy metal resistant bacterium. *Mol Biol Rep* 2020;47:3973–3985.
82. Klonowska A, Moulin L, Ardley JK, Braun F, Gollagher MM, et al. Novel heavy metal resistance gene clusters are present in the genome of *Cupriavidus neocaledonicus* STM 6070, a new species of *Mimosa pudica* microsymbiont isolated from heavy-metal-rich mining site soil. *BMC Genomics* 2020;21:214.
83. Radford DS, Kihlken MA, Borrelly GPM, Harwood CR, Le Brun NE, et al. CopZ from *Bacillus subtilis* interacts *in vivo* with a copper exporting CPx-type ATPase CopA. *FEMS Microbiol Lett* 2003;220:105–112.
84. Kihlken MA, Singleton C, Le Brun NE. Distinct characteristics of Ag⁺ and Cd²⁺ binding to CopZ from *Bacillus subtilis*. *J Biol Inorg Chem* 2008;13:1011–1023.
85. Bhatt K, Timsit E, Rawlyk N, Potter A, Liljebjelke K. Integrative conjugative element ICEHs1 encodes for antimicrobial resistance and metal tolerance in *Histophilus somni*. *Front Vet Sci* 2018;5:153.
86. Cech GM, Szalewska-Patasz A, Kubiak K, Malabirade A, Grange W, et al. The *Escherichia coli* Hfq protein: an unattended DNA-transactions regulator. *Front Mol Biosci* 2016;3:36.
87. Horii T, Ogawa T, Ogawa H. Organization of the recA gene of *Escherichia coli*. *Proc Natl Acad Sci U S A* 1980;77:313–317.
88. Buchmeier NA, Lipps CJ, So MY, Heffron F. Recombination-deficient mutants of *Salmonella typhimurium* are avirulent and sensitive to the oxidative burst of macrophages. *Mol Microbiol* 1993;7:933–936.
89. De Las Peñas A, Connolly L, Gross CA. SigmaE is an essential sigma factor in *Escherichia coli*. *J Bacteriol* 1997;179:6862–6864.
90. Flores-Kim J, Darwin AJ. Regulation of bacterial virulence gene expression by cell envelope stress responses. *Virulence* 2014;5:835–851.
91. Roncarati D, Scarlato V. Regulation of heat-shock genes in bacteria: from signal sensing to gene expression output. *FEMS Microbiol Rev* 2017;41:549–574.
92. Humphreys S, Stevenson A, Bacon A, Weinhardt AB, Roberts M. The alternative sigma factor, sigmaE, is critically important for the virulence of *Salmonella typhimurium*. *Infect Immun* 1999;67:1560–1568.
93. Alghoribi MF, Gibreel TM, Dodgson AR, Beatson SA, Upton M. *Galleria mellonella* infection model demonstrates high lethality of ST69 and ST127 uropathogenic *E. coli*. *PLoS One* 2014;9:e101547.
94. Andrea A, Krogfelt KA, Jenssen H. Methods and challenges of using the greater wax moth (*Galleria mellonella*) as a model organism in antimicrobial compound discovery. *Microorganisms* 2019;7:E85.
95. Jarrell KF, Kropinski AM. The virulence of protease and cell surface mutants of *Pseudomonas aeruginosa* for the larvae of *Galleria mellonella*. *J Invertebr Pathol* 1982;39:395–400.
96. Seed KD, Dennis JJ. Development of *Galleria mellonella* as an alternative infection model for the *Burkholderia cepacia* complex. *Infect Immun* 2008;76:1267–1275.
97. Loutet SA, Valvano MA. A decade of *Burkholderia cenocepacia* virulence determinant research. *Infect Immun* 2010;78:4088–4100.
98. Reasoner DJ, Geldreich EE. A new medium for the enumeration and subculture of bacteria from potable water. *Appl Environ Microbiol* 1985;49:1–7.
99. Kotay SM, Parikh HI, Barry K, Gweon HS, Guilford W, et al. Nutrients influence the dynamics of *Klebsiella pneumoniae* carbapenemase producing enterobacteriales in transplanted hospital sinks. *Water Res* 2020;176:115707.
100. Garratt I, Aranega-Bou P, Sutton JM, Moore G, Wand ME, et al. Long-term exposure to octenidine in a simulated sink trap environment results in selection of *Pseudomonas aeruginosa*, *Citrobacter*, and *Enterobacter* isolates with mutations in efflux pump regulators. *Appl Environ Microbiol* 2021;87:e00210-00221.
101. Wong WC, Dudinsky LA, Garcia VM, Ott CM, Castro VA. Efficacy of various chemical disinfectants on biofilms formed in spacecraft potable water system components. *Biofouling* 2010;26:583–586.
102. Bhagirath AY, Li Y, Somayajula D, Dadashi M, Badr S, et al. Cystic fibrosis lung environment and *Pseudomonas aeruginosa* infection. *BMC Pulm Med* 2016;16:174.

Five reasons to publish your next article with a Microbiology Society journal

1. The Microbiology Society is a not-for-profit organization.
2. We offer fast and rigorous peer review – average time to first decision is 4–6 weeks.
3. Our journals have a global readership with subscriptions held in research institutions around the world.
4. 80% of our authors rate our submission process as 'excellent' or 'very good'.
5. Your article will be published on an interactive journal platform with advanced metrics.

Find out more and submit your article at microbiologyresearch.org.