



PEARL

Evaluation of drugs for potential repurposing against COVID-19 using a tier-based scoring system

Jarvis, Michael A.; Hansen, Frederick A.; Rosenke, Kyle; Haddock, Elaine; Rollinson, Christopher; Rule, Simon; Sewell, Graham; Hughes, Andrew; Feldmann, Heinz

Published in:
Antiviral Therapy

DOI:
[10.3851/imp3368](https://doi.org/10.3851/imp3368)

Publication date:
2020

Link:
[Link to publication in PEARL](#)

Citation for published version (APA):

Jarvis, M. A., Hansen, F. A., Rosenke, K., Haddock, E., Rollinson, C., Rule, S., Sewell, G., Hughes, A., & Feldmann, H. (2020). Evaluation of drugs for potential repurposing against COVID-19 using a tier-based scoring system. *Antiviral Therapy*, 0(0).
<https://doi.org/10.3851/imp3368>

All content in PEARL is protected by copyright law. Author manuscripts are made available in accordance with publisher policies. Wherever possible please cite the published version using the details provided on the item record or document. In the absence of an open licence (e.g. Creative Commons), permissions for further reuse of content should be sought from the publisher or author.

Short communication

Evaluation of drugs for potential repurposing against COVID-19 using a tier-based scoring system

Michael A Jarvis^{1,2*}, Frederick A Hansen³, Kyle Rosenke³, Elaine Haddock³, Christopher Rollinson⁴, Simon Rule⁴, Graham Sewell⁵, Andrew Hughes⁶, Heinz Feldmann³

¹University of Plymouth, Plymouth, Devon, UK

²The Vaccine Group, Ltd, Plymouth, Devon, UK

³Laboratory of Virology, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Hamilton, MT, USA

⁴University Hospitals Plymouth NHS Trust, Plymouth, Devon, UK

⁵The Leicester School of Pharmacy, De Montfort University, Leicester, UK

⁶Manchester Cancer Research Centre, University of Manchester, Manchester, UK

*Corresponding author e-mail: michael.jarvis@plymouth.ac.uk

Background: As the Coronavirus Disease 2019 (COVID-19) pandemic grows daily, we remain with no prophylactic and only minimal therapeutic interventions to prevent or ameliorate severe acute respiratory syndrome Coronavirus 2 (SARS-CoV-2). Prior to SARS-CoV-2 emergence, high throughput screens utilizing clinically developed drugs identified compounds with *in vitro* inhibitory effect on human coronaviruses that may have potential for repurposing as treatment options for COVID-19. However, caution should be applied to repurposing of these drugs when they are taken out of context of human pharmacokinetic parameters associated with normal therapeutic use.

Methods: Our aim was to provide a tier-based scoring system to interrogate this data set and match each drug with its human pharmacokinetic criteria, such as route of administration, therapeutic plasma levels and half-life, tissue distribution, and safety.

Results: Our analysis excluded most previously identified drugs but identified members of 4 drug classes (antimalarial amino-quinolones, selective estrogen receptor modulators; SERMs, low potency tricyclic antipsychotics and tricyclic antidepressants) as potential drug candidates for COVID-19. Two of them, the tricyclic antipsychotics and tricyclic antidepressants were further excluded based on a high adverse event profile.

Conclusions: In summary, our findings using a new pharmacokinetic-based scoring system supports efficacy testing of only a minority of candidates against SARS-CoV-2 infection.

Currently, there is no licensed treatment for COVID-19 worldwide, but the FDA has recently given emergency approval for the use of Remdesivir in COVID-19 patients [1]. In addition, over 1000 clinical trials are currently ongoing or in set-up mode in different countries, including drugs such as lopinavir/ritonavir, dexamethasone, hydroxychloroquine and inhaled interferon beta-1a [2]. Overall, the current situation is far less than satisfying and there is an urgent need for additional treatment options, especially as the pandemic moves into lower resourced countries.

Over the past decade, several studies have used high throughput screens (HTS) to identify clinically developed drugs with *in vitro* inhibitory capacity against human coronaviruses (hCoVs) that may have potential for repurposing as prophylactic or therapeutic treatment options for hCoV infections. These HTS have identified >60 drugs with inhibitory effect as measured by reduction of replication of multiple hCoVs in a variety of different mammalian cell types *in vitro*. However, candidates are rarely considered in light of their pharmacokinetic parameters associated with normal therapeutic dosing. The aim of our study was to use a tier-based scoring system to interrogate this data set by matching drugs with their respective human pharmacokinetic criteria as well as their safety and systemic side effects relevant within the COVID-19 patient setting, allowing us to exclude identified HTS candidates based on these defined pharmacokinetic criteria. Remaining candidates were then further considered based on potential for adverse effects within the COVID-19 patient treatment environment.

Screening clinically approved pharmaceuticals for repurposing removes the substantial time burden associated with movement of experimental drugs from preclinical stage through the regulatory pathway to approval. Repurposing can be especially important for the rapid identification of candidate drugs against emerging infectious diseases such as the present pandemic COVID-19. With notable exceptions [3], only a few drugs have been successfully repurposed, and none for the prevention or treatment of virus infection. With this in mind, our tier-based analysis used three HTS studies as a source of clinically developed drugs with inhibitory effect against *in vitro* replication of multiple hCoVs [4-6]. Drug candidates were critically examined based on the following key pharmacokinetic parameters: route of normal administration, therapeutic plasma levels and half-life, tissue distribution, and safety and adverse reactions. Availability and cost were additional important parameters given the anticipated need for treatment options within low- and middle-income countries. These characteristics were used to i) remove candidates based on pharmacokinetic parameters and potential for adverse events not consistent with prophylactic/therapeutic use for COVID-19, and ii) prioritize remaining drugs for possible *in vitro* confirmation of SARS-CoV-2 inhibitory activity and movement into preclinical animal models (Tier 1 and Tier 2) (Table 1 and Table 2). Tier 1 represents drug candidates with administration, pharmacokinetic and safety parameters suitable for movement into preclinical models; Tier 2 represents similarly suitable candidates, but with a higher adverse event profile. Drugs in Tier 3 have a lower priority due to higher risk of complications in the COVID-19 patient setting, and Tier 4 drugs are those with low prophylactic/therapeutic potential against COVID-19 and/or with high potential for adverse effects (Supplemental Table 1).

In the study of de Wilde *et al* (2014) [5], a 348 FDA-approved drug library was screened for inhibitory activity against Middle East respiratory syndrome CoV (MERS-CoV), with those identified with high inhibitory effect (EC_{50} at low micro-molar concentrations) being tested further for activity against SARS-CoV-1 and hCoV-229E. This HTS resulted in identification of 4 candidates with low micro-molar EC_{50} concentrations against these 3 hCoVs and low cellular toxicity. The Dyall *et al* (2014) study [6] screened by HTS a library of 290 FDA-approved, or experimental drugs with defined molecular targets, that had previously shown activity against RNA viruses [7,8]. The study identified 27 compounds with inhibitory activity against MERS-CoV and SARS-CoV-1 with EC_{50} levels in the low micro-molar range with minimal cytotoxicity. The 2019 HTS of Shen *et al* (2019) [4] screened a 2,000-component library of FDA-approved and pharmacologically active compounds. Seven compounds

were identified with an EC₅₀ of <5 μM against 4 distinct hCoVs. For the purpose of our study, we expanded the inclusion criteria of Shen *et al* (2019) [4] to include a total of 36 compounds with an EC₅₀ of <20μM for the four hCoVs and low cytotoxicity. Together, our analysis was comprised of a total of 58 compounds. We excluded 26 compounds, and identified 11 and 21 compounds with high and medium priority, respectively, with potential for therapeutic intervention against COVID-19. In addition to the single HIV protease inhibitor, lopinovir, the high priority (Tier 1 and Tier 2) compounds represented multiple members from four key drug classes: antimalarial quinolones, selective estrogen receptor modulators (SERMs), amine tricyclic antidepressants, and amine tricyclic antipsychotics.

After removal of primarily experimental agents or those with high toxicity, pharmacokinetic parameters of therapeutic plasma levels of normal dosing and plasma half-life were used as an initial measure to assess whether hCoV inhibitors reach levels required for virus inhibition within the patient. For example, loperamide, an antidiarrheal agent identified as an attractive candidate for repurposing in two HTS screens [4,5] has therapeutic plasma levels >3 orders of magnitude lower than its EC₅₀ against any hCoV tested [9]. This assessment resulted in removal of 26 candidates; drugs applied topically were also removed. Although multiple antipsychotics were identified by HTS as broad inhibitors of hCoV replication *in vitro*, only the low potency 1st generation tricyclics reached the necessary therapeutic plasma levels for inclusion (Tier 2), with the more potent later generation tricyclics commonly orders of magnitude below their EC₅₀.

Four distinct but structurally related members of the antimalarial quinolone class displayed inhibitory effects on multiple hCoVs. Normal therapeutic levels of these agents reached plasma levels approximating the identified EC₅₀ in the HTS studies. Lung tissue distribution (when available) was used as a further parameter, wherein lung tissue-specific accumulation was regarded as a positive indicator for potential therapeutic effect. Based on available data, the quinolone compounds have been shown to accumulate at high (~1000-fold) levels in the lung compared to plasma. Consistent with the prophylactic use of quinolones against malaria, most members exhibit a long plasma half-life, with oral administration being the preferred route to prevent toxicity associated with more rapid parenteral routes. Amongst the quinolones, however, amodiaquine and mefloquine were listed in Tier 2, as compared to other members, these drugs are associated with more severe and prolonged adverse reactions (Table 2). Other high priority candidates with therapeutic plasma levels approaching necessary EC₅₀ levels were the selective estrogen receptor modulators (SERMs) (tamoxifen and toremifene), amine tricyclic antidepressants (clomipramine and desipramine), and low potency tricyclic antipsychotics (promazine and chlorpromazine). The SERMs in particular showed high accumulation within the lung and minimal adverse reactions, and were listed in Tier 1 (Table 1). In contrast, due to a higher number of associated adverse reactions, as well as the frequent need for optimization of the dosing regimen, the antidepressants and antipsychotics were listed in Tier 2 (Table 2) [9-11].

Multiple hCoV inhibitory drugs identified by HTS with attractive pharmacokinetic profiles were prioritized lower (Tier 3) due to either their preclinical/experimental status or higher possibility for adverse reactions in the COVID-19 patient setting; otherwise attractive candidates with short half-lives were also included in this tier (Supplemental Table 1). Several antineoplastic agents were identified that interfere with DNA and RNA replication. The prodrug mycophenolate mofetil, through its mycophenolic acid active metabolite, is an inhibitor of guanosine synthesis through an inhibitory effect on

inosine monophosphate dehydrogenase; gemcitabine and hycanthone both inhibit DNA and RNA synthesis directly through distinct mechanisms. Depending on the dosage, these anti-neoplastic drugs can result in a level of immune suppression that may be contraindicated for use in COVID-19 patients, where adaptive immunity will presumably be important. Dasatinib and imatinib, small molecule inhibitors of the Abl tyrosine kinase pathway, also fell within Tier 3 for this reason. Metronomic dosing to achieve plasma levels above the EC_{50} , but below the immunosuppressive dose may achieve the necessary level of virus inhibition without undermining patient adaptive immune responses. Such a treatment scenario would need to be assessed in a preclinical animal model before moving to clinical studies.

Timing of therapeutic intervention against SARS-CoV-2 appears to be critical, with disease etiology changing over time: disease being a more direct effect of virus replication at early times, whilst later lung pathology being host immune response-driven. The HTS studies detailed above identify compounds based on inhibition of hCoV replication. *Drug candidates are therefore expected to be more effective for COVID-19 disease management when used early.* Studies using Remdesivir (GS-5734; hCoV polymerase inhibitor) highlight the altering course of disease over time and the importance of instigating antiviral measures early. Later drug administration reduced virus replication, but failed to improve lung function or disease outcome in the immunopathologic-driven stage of disease [12]. This key importance of timing may be a possible explanation for inconsistent results from recent and ongoing studies investigating repurposing of drugs such as hydroxychloroquine, which again emphasizes the need for preclinical animal challenge models before progression to human clinical trials.

To be useful clinically, drugs will need to have a minimal adverse event profile (particularly for prophylaxis); not be contraindicated in patients who have underlying medical conditions; and achieve therapeutic drug concentrations rapidly. The low potency tricyclic antipsychotics do not possess these attributes; both chlorpromazine and promazine need to be carefully titrated to optimal therapeutic doses, and many patients report a plethora of adverse events. Furthermore, they are contraindicated in patients with multiple co-morbidities which place these patients into the COVID-19 vulnerable category. The amine tricyclic anti-depressants clomipramine and desipramine face similar challenges. Half of treated patients may report somnolence and dizziness aside of other adverse events. Patients with underlying medical conditions are either more likely to experience some of these class toxicities (e.g. glaucoma, urinary retention from their anti-cholinergic properties) or be contraindicated. Furthermore, they show potential for overdose misuse and suicidal ideation as well as withdrawal symptoms even after short courses of treatment.

The protease inhibitor lopinavir, the SERMS tamoxifen and toremifene and the anti-malarials chloroquine and hydroxychloroquine have wider therapeutic indices than the tricyclic drugs and have decades of widespread clinical use across geographies, patient demographics and co-morbidities. Lopinavir's recommended daily dose for HIV-1 infection (800mg) produces plasma levels covering the EC_{50} values for pathogenic hCoVs. Tamoxifen and toremifene are customarily used at daily doses of 20mg and 60mg, respectively, for breast cancer. But higher daily doses (~600mg and 680mg, respectively) are relatively well tolerated under short durations reaching plasma concentrations after a single dose at anti-viral EC_{50} levels and evidence of greater concentrations within tissue.

COVID-19 is a rapidly evolving situation. During review of the manuscript some of the drugs under consideration were tested for *in vitro* inhibitory activity against

SARS-CoV-2, which is shown in the accompanying tables (Table 1 & 2 and Supplemental Table 1). During this time, two pre-clinical animal studies have also reported the absence of any effect of hydroxychloroquine when used either prophylactically or therapeutically against SARS-CoV-2 replication or associated disease [13,14]. Hydroxychloroquine treatment was also recently removed from the WHO Solidarity COVID-19 clinical study based on evidence from the Solidarity trial, a Cochrane review of the drug as well as on the release of a report from the UK-based RECOVERY trial where hydroxychloroquine showed no effect on mortality rate of COVID-19 patients [15,16]. Similarly, a post-exposure prophylaxis trial showed no effect of hydroxychloroquine on the incidence of infection from high and moderate-risk exposure to SARS-CoV-2 [17]. Multiple pre-exposure prophylaxis trials remain ongoing [18]. New recent data has also shed light on a possible mechanism behind the apparent divergence in inhibitory effect of hydroxychloroquine between *in vitro* and *in vivo* studies; wherein, the virus uses a distinct entry pathway in the Vero cells standardly used for *in vitro* determination of drug sensitivity, compared to the pathway utilized in lung epithelium *in vitro* and presumably *in vivo*. Notably, only the entry pathway in Vero cells is susceptible to inhibition by endosomal pathway inhibitors such as hydroxychloroquine [19,20].

In summary, caution should be applied to repurposing of drugs when they are taken out of context of human pharmacokinetic parameters associated with normal therapeutic use. Our tier-based scoring system to analyze drugs identified through HTS with *in vitro* efficacy against one or more hCoVs resulted in the exclusion of the majority of compounds for further consideration. Similar to the quinolones, SERMs (i.e. tamoxifen and toremifene) are a class of drugs that have characteristics of low micro-molar hCoV inhibitory activity, attractive human pharmacokinetics, favorable tissue accumulation and good safety profile for use in COVID patients [9]. The next step for all potential candidates will be preclinical efficacy testing in animal models against SARS-CoV-2 challenge. Repurposing of clinically approved drugs helps remove the concern of overt drug toxicity. However, animal infection models are critical as they place the treatment within the context of the kinetics of virus infection within the host. They can also identify unexpected enhancement of disease by a drug in context of viral infection, as experienced with mycophenolate mofetil against MERS-CoV in nonhuman primates [21]. A similar enhancement effect was seen for chloroquine prophylaxis but not treatment of mosquito-transmitted chikungunya, which corresponded with an immunomodulatory effect of the drug, and again emphasizes the importance of timing in therapeutic intervention [22].

Finally, combinations of drugs are often far more effective than single compounds [23]. Therefore, these Tier 1 drugs should be considered for combined use to take advantage of possible synergy between drugs with differing modalities of virus inhibition. However, unpredicted antagonism can also result from such combinations, as was recently observed between chloroquine and Remdesivir [24]. Again, such studies initially need to be performed using *in vitro* cell systems and, importantly, preclinical animal models prior to considering movement into humans.

Acknowledgements

This work was partially funded through awards to the Vaccine Group Company, Ltd, and the University of Plymouth; and was partially funded by the Intramural Research Program of NIAID, NIH.

Disclosure Statement

AH is a former employee of AstraZeneca and current shareholder of AstraZeneca who licenses tamoxifen. Other authors claim no conflict of interest.

References

1. FDA. Drug Approvals and Database 2020. (Accessed 22 July 2020). Available from <https://www.fda.gov/drugs/development-approval-process-drugs/drug-approvals-and-databases>.
2. WHO. COVID-19: International Clinical Trials Registry Platform (ICTRP) 2020. (Accessed 22 July 2020). Available from <https://www.who.int/ictcp/search/en/>.
3. Azvolinsky A. Repurposing Existing Drugs for New Indications. *The Scientist* [Internet]. 2016 (Accessed 22 July 2020). Available from <https://www.the-scientist.com/features/repurposing-existing-drugs-for-new-indications-32285>.
4. Shen L, Niu J, Wang C, Huang B, *et al*. High-Throughput Screening and Identification of Potent Broad-Spectrum Inhibitors of Coronaviruses. *J Virol* 2019; **93**:e00023-e00019
5. de Wilde AH, Jochmans D, Posthuma CC, *et al*. Screening of an FDA-approved compound library identifies four small-molecule inhibitors of Middle East respiratory syndrome coronavirus replication in cell culture. *Antimicrob Agents Chemother* 2014; **58**:4875-84.
6. Dyall J, Coleman CM, Hart BJ, *et al*. Repurposing of clinically developed drugs for treatment of Middle East respiratory syndrome coronavirus infection. *Antimicrob Agents Chemother* 2014; **58**:4885-4893.
7. Lehar J, Krueger AS, Avery W, Heilbut AM, *et al*. Synergistic drug combinations tend to improve therapeutically relevant selectivity. *Nat Biotechnol* 2009; **27**:659-666.
8. Johansen LM, Brannan JM, Delos SE, *et al*. FDA-approved selective estrogen receptor modulators inhibit Ebola virus infection. *Sci Transl Med* 2013; **5**:190ra79.
9. AHFS. AHFS Drug Information 2020.
10. Winstanley P, Edwards G, Orme M, Breckenridge A. The disposition of amodiaquine in man after oral administration. *Br J Clin Pharmacol* 1987; **23**:1-7.
11. Brunton L, Knollmann B, Hilal-Dandan R. Goodman and Gilman's The Pharmacological Basis of Therapeutics. 13th ed: McGraw-Hill Education; 2017.
12. Sheahan TP, Sims AC, Graham RL, *et al*. Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses. *Sci Transl Med* 2017; **9**:aal3653.
13. Maisonnasse P, Guedj J, Contreras V, *et al*. Hydroxychloroquine in the treatment and prophylaxis of SARS-CoV-2 infection in non-human primates. *Research Square* 2020. Epub 2020/05/06. doi: 10.21203/rs.3.rs-27223/v1.
14. Rosenke K, Jarvis MA, Feldmann F, *et al*. Hydroxychloroquine Proves Ineffective in Hamsters and Macaques Infected with SARS-CoV-2. *bioRxiv*. 2020. Epub 2020/06/25. doi: 10.1101/2020.06.10.145144.
15. WHO. "Solidarity" clinical trial for COVID-19 treatments 2020. (Accessed 22 July 2020). Available from <https://www.who.int/emergencies/diseases/novel-coronavirus-2019/global-research-on-novel-coronavirus-2019-ncov/solidarity-clinical-trial-for-covid-19-treatments>.
16. No clinical benefit from use of hydroxychloroquine in hospitalised patients with COVID-19 [Internet]. 2020; 5 June, 2020. (Accessed 22 July 2020). Available from <https://www.recoverytrial.net/news/statement-from-the-chief>

[investigators-of-the-randomised-evaluation-of-covid-19-therapy-recovery-trial-on-hydroxychloroquine-5-june-2020-no-clinical-benefit-from-use-of-hydroxychloroquine-in-hospitalised-patients-with-covid-19](#)

17. Boulware DR, Pullen MF, Bangdiwala AS, *et al.* A Randomized Trial of Hydroxychloroquine as Postexposure Prophylaxis for Covid-19. *N Engl J Med* 2020. Epub 2020/06/04.
18. National Institutes of Health UNLoM. Clinicaltrials.gov 2020. (Accessed 22 July 2020). Available from <https://clinicaltrials.gov>.
19. Yamamoto M, Kiso M, Sakai-Tagawa Y, Iwatsuki-Horimoto K, Imai M, Takeda M, *et al.* The Anticoagulant Nafamostat Potently Inhibits SARS-CoV-2 S Protein-Mediated Fusion in a Cell Fusion Assay System and Viral Infection In Vitro in a Cell-Type-Dependent Manner. *Viruses* 2020; **12**:E629
20. Dittmar M, Lee JS, Whig K, Segrist E, Li M, Jurado K, *et al.* Drug repurposing screens reveal FDA approved drugs active against SARS-Cov-2. *bioRxiv*. 2020. Epub 2020/06/19. doi: 10.1101/2020.06.19.161042.
21. Chan JF, Yao Y, Yeung ML, *et al.* Treatment With Lopinavir/Ritonavir or Interferon-beta1b Improves Outcome of MERS-CoV Infection in a Nonhuman Primate Model of Common Marmoset. *J Infect Dis* 2015; **212**:1904-1913.
22. Roques P, Thiberville SD, Dupuis-Maguiraga L, *et al.* Paradoxical Effect of Chloroquine Treatment in Enhancing Chikungunya Virus Infection. *Viruses* 2018; **10**:268
23. Maeda K, Das D, Kobayakawa T, Tamamura H, Takeuchi H. Discovery and Development of Anti-HIV Therapeutic Agents: Progress Towards Improved HIV Medication. *Curr Top Med Chem* 2019; **19**:1621-1649.
24. FDA. Remdesivir by Gilead Sciences: FDA Warns of Newly Discovered Potential Drug Interaction That May Reduce Effectiveness of Treatment <https://www.fda.gov/safety/medical-product-safety-information/remdesivir-gilead-sciences-fda-warns-newly-discovered-potential-drug-interaction-may-reduce>.
25. Regenthal R, Krueger M, Koepfel C, Preiss R. Drug levels: therapeutic and toxic serum/plasma concentrations of common drugs. *J Clin Monit Comput* 1999; **15**:529-544.
26. Durcan L, Clarke WA, Magder LS, Petri M. Hydroxychloroquine Blood Levels in Systemic Lupus Erythematosus: Clarifying Dosing Controversies and Improving Adherence. *J. Rheumatology* 2015; **42**:2092-2097
27. McChesney EW, Banks WF, Fabian RJ. Tissue Distribution of Chloroquine, Hydroxychloroquine, and Desethylchloroquine in the Rat. *Toxicology and Applied Pharmacology* 1967; **10**:501-513.
28. Weston S, Haupt R, Logue J, Matthews K, Frieman M. FDA approved drugs with broad anti-coronaviral activity inhibit SARS-CoV-2 *in vitro*. *bioRxiv* 2020; 10.1101/2020.03.25.008482
29. Lien EA, Solheim E and Ueland PM. Distribution of Tamoxifen and Its Metabolites in Rat and Human Tissues during Steady-State Treatment. *Cancer Res* 1991; **51**:4837-4844
30. Martei YM, Pace LE, Brock JE, Shulman LN. Breast Cancer in Low- and Middle-Income Countries: Why We Need Pathology Capability to Solve This Challenge. *Clin Lab Med* 2018; **38**:161-173

31. Liippo K, Ellmen J, Vanttinen E, Anttila M. Toremifene concentration and multidrug resistance in lung tumors. *Cancer Chemother Pharmacol* 1997; **39**:212-216
32. Atzori C, Villani P, Regazzi M, Maruzzi M, Cargnel A. Detection of intrapulmonary concentration of lopinavir in an HIV-infected patient. *AIDS* 2003; **17**:1710-1711.
33. Potts AM. The concentration of phenothiazines in the eye of experimental animals. *Invest Ophthalmol* 1962; **1**:522-530.

Supplementary Table 1		Tier 3				
		Name of Drug (mol. wt. g/mol)	Plasma Levels & Lung Distribution	Half-life	EC ₅₀ , CC ₅₀ (μM)	Reason for Tier Designation
Antineoplastic Drugs	Kinase signaling inhibitor	Dasatinib¹ (488.0)	Oral administration of 100mg results in 104ng/ml (0.2uM). ¹⁰ Accumulates (15-fold) in lung compared to plasma. ¹¹	< 4 h ¹⁰	MERS(5.468); SARS(2.100) ¹	Pros: 1) Lung levels within range of EC ₅₀ 2) Good clinical experience Cons: 1) Short half-life 2) Potential immune suppression (potential for metronomic dosing)
		Imatinib mesylate¹ (589.7)	Following a 400mg oral dose given once daily to steady state peak plasma concentration of 1.8ug/ml (3.05uM) to 3.4ug/ml (5.76uM). ⁴ Lung accumulation is not known.	22h ⁴	MERS(17.689); SARS(9.823) ¹ ; SARS-CoV-2(5.32, >30.86) ⁴⁰	Pros: 1) Lung levels within range of EC ₅₀ 2) Good clinical experience Cons: 1) Short half-life 2) Potential immune suppression (potential for metronomic dosing)
	Ribonucleotide reductase inhibitor	Gemcitabine HCl¹ (299.7)	Steady-state concentration during a 10mg/m ² per min infusion for 120 to 640 min is 26.9uM. Used for non-small cell lung cancer indicating clinically significant lung localization. ⁴	15 min to 1 h ⁴	MERS(1.216); SARS(4.957) ¹	Pros: 1) Lung levels within range of EC ₅₀ 2) Good clinical experience Cons: 1) Short half-life 2) Potential immune suppression (potential for metronomic dosing)
	DNA/ RNA synthesis inhibitor	Hycanthone⁸ (356.5)	Infusion of 60mg/m ² results in two patients resulted in peak plasma of 1ug/ml (2.80uM) and following 100mg/m ² a peak plasma of 2.4ug/ml (6.73uM). ¹² Lung distribution not known.	3 to 5 hours ¹²	OC43(0.16, 3.58); NL63(5.76, 3.68); MERS(5.11, 4.32); A59(5.78, 4.19) ⁸	Pros: 1) Presume lung levels within range of EC ₅₀ Cons: 1) Short half-life 2) Potential immune suppression (potential for metronomic dosing) 3) No clinical experience
Immunosuppressants	Guanosine synthesis inhibitors	Mycophenolate mofetil⁸ (433.5)	Prodrug rapidly converted to active drug MPA (see below).	See Below	OC43(1.58, 3.43); NL63(0.23, 3.01); MERS(1.54, 3.17); A59(0.27, 3.33) ⁸ ; SARS-CoV-2(0.47, >10) ⁴¹	Pros: 1) Lung levels within range of EC ₅₀ 2) Good clinical experience 3) Suitable half-life Cons: 1) Potential immune suppression (potential for metronomic dosing)
		Mycophenolic acid (MPA)⁸ (320.3)	8ug/ml (24.97uM) to 19ug/ml (59.32uM) following a 1 to 1.75 g oral dose given twice daily to steady state in renal patients. ⁴	11 to 24 h ⁴	OC43(1.95, 3.55); NL63(0.18, 3.44); MERS(1.95, 3.21); A59(0.17, 4.18) ⁸	Pros: 1) Lung levels within range of EC ₅₀ 2) Good clinical experience 3) Suitable half-life Cons: 1) Potential immune suppression (potential for metronomic dosing)
Antihypertensives	Beta blocker	Alprenolol⁸ (249.3)	Therapeutic levels 0.025ug/ml (0.10uM) to 0.14ug/ml (0.56uM). ⁶ High lung accumulation after IV administration, but only 2-fold after oral administration. ¹³	2 to 7 h ⁶	OC43(1.95, >20); NL63(11.88, >20); MERS(10.53, >20); A59(13.97, >20) ⁸	Pros: 1) Lung levels within range of EC ₅₀ 2) High lung accumulation and be achieved (IV administration) 3) Good clinical experience Cons: 1) Short half-life
		Propranolol⁸ (259.3)	Therapeutic level 0.02ug/ml (0.07uM) to 0.3ug/ml (1.15uM). ⁶ Rabbits given a 10mg/kg subcutaneous dose had 250-fold higher levels of drug in lung than blood at 1h. ¹⁴ In dogs given a 4.5mg/kg dose over 45 min, 50-fold higher levels were present in lung compared to plasma. ¹⁵	2 to 6 h ⁶	OC43(0.48, >20); NL63(8.11, >20); MERS(11.01, >20); A59(13.54, >20) ⁸	Pros: 1) Lung levels within range of EC ₅₀ 2) High lung accumulation 3) Good clinical experience Cons: 1) Short half-life
	Alpha-1 adrenergic blocker	Doxazosin mesylate⁸ (547.6)	0.01ug/ml (.018uM) to 0.15ug/ml (0.27uM) ⁶ Accumulation data not available, but prototype of family, prazosin, accumulates in lungs. ¹⁶	20h ⁴	OC43(4.97, >20); NL63(13.95, >20); MERS(12.66, >20); A59(14.48, >20) ⁸	Pros: 1) At lower limit of EC ₅₀ 2) Parent, prazosin, accumulates 10-fold in lungs Cons: 1) At lower limit of EC ₅₀

	Vasodilator/ Muscle relaxant	Papaverine⁸ (339.4)	Based on 80mg oral dose, plasma levels of 0.049ug/ml (0.14uM) to 0.314ug/ml (0.93uM). ¹⁷ Normal adult dose is 150mg every 8-12 h. Localizes to liver and fat deposits. Lung distribution not known. ¹⁸	3 h ¹⁷	OC43(1.61, 12.11); NL63(7.32, 11.71); MERS(9.45, 11.98); A59(11.46, 12.44) ⁸	Pros: 1) Levels at low range of EC ₅₀ 2) Good clinical experience Cons: 1) Levels at low range of EC ₅₀ 2) Short half life 3) Lung accumulation unknown
Neurotransmitter Inhibitors	Antihistamines	Astemizole¹ (458.6)	0.002ug/ml (0.0045uM) to 0.05ug/ml (0.11uM). ⁶ High lung accumulation of both forms. Daily oral dosing of dogs with 1mg/kg for 6 weeks resulted in 725 to 1020-fold of AST and DES-AST combined over plasma level. ¹⁹	20 to 26 h (Astemizole) 9 days (Desmethyl-astemizole) ⁶	MERS(4.884); SARS(5.591) ¹	Pros: 1) Lung levels within range of EC ₅₀ 2) High lung accumulation 3) Good clinical experience Cons: 1) Short half-life 2) Withdrawn due to rare fatal arrhythmias
		Chlorphenoxamine HCl¹ (340.3)	After 40mg oral dose, plasma levels below 10ng/ml (0.03uM) (limit of detection). Based on diphenhydramine, lack of significant lung uptake. ²⁰	2-9 h (diphenhydramine analog) ⁴	MERS(12.646); SARS(20.031) ¹	Pros: 1) Good clinical experience Cons: 1) Levels far below EC ₅₀ 2) short half-life 3) assume poor lung accumulation based on diphenhydramine
		Chloropyramine⁸ (289.8)	Not approved for use in US. In dogs, 7.5mg/ml oral administration results in peak plasma levels of 234ng/ml (0.8uM). ²¹ Lung approximately 80-fold higher than plasma level after 6h in mice. ²²	21 h ²¹	OC43(1.79, >20); NL6(14.21, >20); MERS(14.21, >20); A59(2.42, >20) ⁸	Pros: 1) Lung levels within range of EC ₅₀ 2) High lung accumulation 3) Suitable half-life Cons: 1) Not approved for use in humans
		Conessine⁸ (356.6)	No pharmacokinetic data available. Based on a N-demethylate derivative (DMNG-3) in rats, an oral 40mg/kg dose resulted in 2.06ug/ml (5.78uM); 20mg/kg dose 0.97ug/ml (2.72uM). ²³ After oral administration to mice distributes to all tissues but accumulates in GI and liver, with moderate to low accumulation in lungs. ²³	15 h ²³	OC43(2.34, >20); NL63(10.75, >20); MERS(4.98, >20); A59(11.46, >20) ⁸	Cons 1) Experimental 2) Moderate to low accumulation in lungs
		Promethazine HCl¹ (320.9)	0.05ug/ml (0.16uM) to 0.4ug/ml (1.2uM) ⁶	8 to 15 h ⁶	MERS(11.802); SARS(7.545) ¹ ; SARS-CoV-2(10.44, >42.59) ⁴⁰	Pros: 1) Levels at low range of EC ₅₀ 2) Good clinical experience Cons: 1) Levels at low range of EC ₅₀ 2) Lung accumulation unknown
	Anticholinergic	Benztropine mesylate¹ (403.5)	1.5mg single oral dose 2.5ng/ml (0.006uM) ²⁴	7 h ²⁴	MERS(16.627); SARS(21.611) ¹ ; SARS-CoV-2(17.79, >50) ⁴⁰	Pros: 1) Good clinical experience Cons 1) Levels far below EC ₅₀ 2) short half-life 3) lung accumulation unknown
Antiparasitics	Anti-amoebic	Emetine^{1,8} (553.6)	Peak plasma levels following oral ipecac is 9.6ug/ml (0.02uM). ²⁵ High lung accumulation ²⁶	24 to 48 h ⁶	OC43(0.30, 2.69); NL63(1.43, 3.63); MERS(0.34, 3.08); A59(0.12, 3.51) ⁸ MERS (0.014); SARS (0.051) ¹ ; SARS-CoV-2(<0.01, .7) ⁴²	Pros: High lung accumulation 3) Good clinical experience 4) Long half-life Cons: 1) Lung levels below range of EC ₅₀ (may be overcome by lung accumulation) 2) Induces emesis in patients
	Protein synthesis inhibitor	Lycorine⁸ (287.3)	Peak plasma concentration in mice 5.1ug/ml (17.75uM). Distributes widely to tissues, including lungs, but then reduced to undetectable levels over 2 h. ²⁷	3 to 6 h ²⁷	OC43(0.15, 4.37); NL63(0.47, 3.81); MERS(1.63, 3.14); A59(0.31, 3.51) ⁸ ; SARS-CoV-2(0.31, >40) ⁴³	Cons: 1) Experimental 2) Short half-life

Other	Calcium channel blocker	Tetrandrine⁸ (622.8)	Oral administration of 100mg results in 67ng/ml (0.1uM) peak plasma concentration. Berbamine is a key metabolite, with same study showing 33ng/ml (0.05uM). ²⁸ Concentrated 8-fold in lungs compared to plasma. ²⁹	24hr ²⁸	OC43(.29, >20); NL63(2.05, >20); MERS(12.68, >20); A59(4.81, >20) ⁸	Cons: 1) Experimental in US. Approved in China for treatment of silicosis.
	Interferon inducer	Tilorone⁸ (483.5)	At a well tolerated 10mg/kg dose, peak plasma concentration is 135ng/ml (0.28uM) in males and 92.3ng/ml (0.19uM) in females. At 2mg/kg, 50.5ng/ml (0.10uM) and 17.5ng/ml (0.036uM), males and females, respectively. ³⁰ In mice 20-fold higher accumulation in lung than serum. ³¹	20 h ³⁰	OC43(0.32, >20); NL63(6.89, >20); MERS(10.56, >20); A59(16.11, >20) ⁸	Cons: 1) Experimental in US. Marketed as Amixin, Lavomax as an antiviral in Russia
	Antiplatelet	Ticlopidine⁸ (263.8)	For reduction of risk nonfatal stroke normal dose is 250mg twice daily. Oral administration of 250mg results in peak plasma concentration of 0.08ug/ml (0.30uM) to 0.8ug/ml (3.03uM). ³² Lung distribution not known.	20 to 50 h ³²	OC43(1.41, >20); NL63(15.65, >20); MERS(11.25, >20); A59(14.28, >20) ⁸	Pros: Within range of EC ₅₀ 2) Good clinical experience 3) Long half-life Cons: 1) Lung accumulation unknown 2) Intensive patient management
	Low potency tricyclic antipsychotics	Triflupromazine HCl¹ (388.9)	No PK data. Assume comparable to chlorpromazine of this class.	23 to 37 h ⁴	MERS(5.758); SARS(6.398) ¹	Pros: 1) Comparable to Tier 1 low potency antipsychotics Cons: 1) Discontinued in US
Tier 4						
Antimicrobials	Antibacterials	Anisomycin¹ (265.3)	NR*	NR	MERS(0.003); SARS(0.191) ¹	Cons: 1) Experimental 2) High toxicity
		Salinomycin sodium⁸ (773.0)	NR	NR	OC43(0.29, 1.97); NL63(5.71, 2.41); MERS(5.49, 3.84); A59(5.16, 2.45) ⁸	Cons: 1) High toxicity 2) Not used in humans. Animal feed additive.
		Valinomycin⁸ (1,111.3)	NR	NR	OC43(4.43, 6.15); NL63(1.89, 4.12); MERS(6.07, 5.88); A59(6.78, 5.11) ⁸	Cons: 1) High toxicity
		Dihydrocelestryl diacetate⁸ (536.7)	NR	NR	OC43(1.17, >20); NL63(0.65, >20); MERS(10.58, >20); A59(4.24, >20) ⁸	Cons: 1) Experimental use
		Cetylpyridinium chloride⁸ (340.0)	NR	NR	OC43(4.31, 8.23); NL63(1.24, 8.52); MERS(0.69, 8.14); A59(7.86, 8.19) ⁸	Cons: 1) Cationic disinfectant 2) Ingredient of mouthwash and poorly absorbed 3) No systemic PK data
		Monensin sodium⁸ (692.9)	NR	NR	OC43(3.81, >20); NL63(1.54, >20); MERS(3.27, >20); A59(0.18, >20) ⁸	Cons: 1) High toxicity 2) Not used in humans.
		Oligomycin⁸ (791.1)	NR	NR	OC43(0.19, 6.56); NL63(2.63, 4.26); MERS(0.21, 5.16); A59(6.43, 6.78) ⁸	Cons: 1) High toxicity
	Antifungals	Cycloheximide^{1,8} (281.4)	NR	NR	OC43(0.43, 3.12); NL63(2.64, 3.24); MERS(2.56, 2.96); A59(5.21, 3.19) ⁸ MERS(0.189); SARS(0.043) ¹ ; SARS-CoV-2(0.58) ⁴⁴	Cons: 1) High toxicity
		Exalamide⁸ (221.3)	NR	NR	OC43(1.48, >20); NL63(17.49, >20); MERS(15.91, >20); A59(16.39, >20) ⁸	Topical application (anti-fungal). Not used systemically.

	Ant	Phenylmercuric acetate ⁸ (336.7)	NR	NR	OC43(2.17, 5.35); NL63(6.79, 5.47); MERS(6.44, 5.39); A59(6.81, 5.97) ⁸	Cons: 1) High toxicity
		Terconazole ¹ (532.5)	Topical preparation	NR	MERS(12.203); SARS (15.327) ¹ ; SARS-CoV-2(16.14, 41.46) ⁴⁰	Cons: Topical antifungal
	Anti-pinworm	Pyruvium pamoate ⁸ (769.9)	Single oral dose of 350mg. Not absorbed after oral administration. ³³	Not absorbed ³³	OC43(3.21, >20); NL63(3.35, >20); MERS(1.84, 19.91); A59(4.12, 19.98) ⁸	Oral anti-helminth Cons: 1) Plasma levels far below EC ₅₀ (not absorbed)
Neurotransmitters	Antipsychotics	Fluphenazine HCl ¹ (510.4)	0.001ug/ml (0.002uM) to 0.004ug/ml(0.008uM) ⁶	8 to 28 h ⁴ (depending on route and formulation)	MERS(5.868); SARS(21.431) ¹ ; SARS-CoV-2(8.98, 20.02) ⁴⁰	Cons: 1) Plasma levels far below EC ₅₀
		Fluspirilene ¹ (475.6)	Single 2mg intramuscular dose resulted in peak plasma of 0.000083ug/ml (0.0001uM) to 0.000280ug/ml (0.0006uM) ³⁴	NR	MERS(7.477); SARS(5.963) ¹ ; SARS-CoV-2(5.32, 30.33) ⁴⁰	Cons: 1) Plasma levels far below EC ₅₀
		Thiothixene ¹ (443.6)	0.001ug/ml (0.002uM) to 0.025ug/ml (0.056uM) ⁶	34 h ⁴	MERS(9.297); SARS(5.316) ¹	Cons: 1) Plasma levels far below EC ₅₀
Anti-hypertensives	Calcium channel blocker	Berbamine ⁸ (608.7)	Oral administration of 100mg results in peak plasma of 33ng/ml (0.05uM). ²⁸ Lung distribution not known.	39 h ²⁸	OC43(1.48, >20); NL63(9.46, >20); MERS(13.14, >20); A59(10.91, >20) ⁸	Cons: 1) Experimental use
	Chalcone	4'-Hydroxychalcone ⁸ (224.3)	Experimental with minimal information. No PK data	NR	OC43(1.52, >20); NL63(7.25, >20); MERS(10.23, >20); A59(9.75, >20) ⁸	Cons: 1) Experimental use
Other	Oxidative phosphorylation inhibitor	Antimycin A ⁸ (548.6)	NR	NR	OC43(1.65, 3.62); NL63(6.05, 4.21); MERS(6.89, 4.32); A59(5.42, 3.98) ⁸	Cons: 1) High toxicity
	Local anaesthetic	Diperodon ⁸ (433.9)	NR	NR	OC43(1.71, 14.3); NL63(4.91, 13.6); MERS(8.77, 14.2); A59(1.98, 14.4) ⁸	Used as topical anaesthetic
	Cathepsin inhibitor	E-64-D ¹ (342.4)	Not known	Not known	MERS(1.275); SARS(0.760) ¹	Cons: 1) Experimental use
	Beta-carboline alkaloid	Harmine ⁸ (212.3)	Neurotoxin	NR	OC43(1.9, >20); NL63(13.46, >20); MERS(4.93, >20); A59(13.77, >20) ⁸	Cons: 1) High toxicity 2) Experimental use
	Anti-diarrheal	Loperamide ^{5,8} (513.5)	Standard oral dose of 2mg results in peak plasma concentration of 0.002ug/ml (0.0039uM). ³ Abuse with toxicity using higher doses can exceed 100ng/ml (0.038uM). Lung accumulation not known. In one overdose study, lung not tested but 5-fold increase in liver over peripheral blood. ³⁵	11 h ⁴	MERS(4.8, 15.5); SARS(5.9, 53.8); 229E(4.0, 25.9) ⁵ OC43(1.86, 18.7); NL63(6.47, 18.27); MERS(4.82, 18.9); A59(10.65, 18.9) ⁸	Cons: 1) High toxicity 2) Levels far below EC ₅₀

	Urinary analgesic	Phenazopyridine⁸ (213.2)	Following standard 200mg oral dose, peak plasma levels of 0.012ug/ml (0.056uM). ³⁶ No substantial accumulation in the lung (maximum 1.8-fold at 2 h post 100mg/kg dose in rats). ³⁷	50 min ³⁶	OC43(1.90, <20); NL63(2.02, >20); MERS(1.93, >20); A59(0.77, >20) ⁸	Cons: 1) At lower limit of EC ₅₀ 2) Does not accumulate in lungs 3) Short half life
	Quinoid-type triterpene	Pristimerin⁸ (464.6)	Following and oral dose of 2mg/kg in rats, peak plasma level 0.19ug/ml (0.41uM). ³⁸ Tissue distribution is not known.	5h ³⁸	OC43(1.99, >20); NL63(1.63, >20); MERS(13.87, >20); A59(9.17, >20) ⁸	Cons: 1) Experimental 2) Levels far below EC ₅₀
	Muscle relaxant	Zoxazolamine⁸ (168.6)	Single oral dose of 0.75 to 1g results in peak plasma levels of 3ug/ml (17.8uM) to 12ug/ml (71.2uM). ³⁹ Lung accumulation not known, but no accumulation in other multiple tissues (muscle, kidney, liver, brain or fat) in dogs. ³⁹	Rapid (completely gone by 7 h ³⁹)	OC4391.39, >20); NL63(13.51, >20); MERS(14.21, >20); A59(16.45, >20) ⁸	Cons: 1) High liver toxicity
NR: Not relevant						

References (Supplemental Table)

1. Dyall J, Coleman CM, Hart BJ, *et al.* Repurposing of clinically developed drugs for treatment of Middle East respiratory syndrome coronavirus infection. *Antimicrob Agents Chemother* 2014; **58**:4885-4893.
2. Winstanley P, Edwards G, Orme M, Breckenridge A. The disposition of amodiaquine in man after oral administration. *Br J Clin Pharmacol* 1987; **23**:1-7.
3. AHFS. AHFS Drug Information 2020.
4. Brunton L, Knollmann B, Hilal-Dandan R. Goodman and Gilman's The Pharmacological Basis of Therapeutics. 13th ed: McGraw-Hill Education; 2017.
5. de Wilde AH, Jochmans D, Posthuma CC, *et al.* Screening of an FDA-approved compound library identifies four small-molecule inhibitors of Middle East respiratory syndrome coronavirus replication in cell culture. *Antimicrob Agents Chemother* 2014; **58**:4875-4884.
6. Regenthal R, Krueger M, Koeppel C, Preiss R. Drug levels: therapeutic and toxic serum/plasma concentrations of common drugs. *J Clin Monit Comput* 1999; **15**:529-544.
7. Potts AM. The concentration of phenothiazines in the eye of experimental animals. *Invest Ophthalmol* 1962; **1**:522-530.
8. Shen L, Niu J, Wang C, *et al.* High-Throughput Screening and Identification of Potent Broad-Spectrum Inhibitors of Coronaviruses. *J Virol* 2019; **93**:e00023-e00019
9. Aitchison K, Datla K, Rooprai H, Fernando J, Dexter D. Regional distribution of clomipramine and desmethylclomipramine in rat brain and peripheral organs on chronic clomipramine administration. *J Psychopharmacol* 2010; **24**:1261-1268.

10. Christopher LJ, Cui D, Wu C, et al. Metabolism and disposition of dasatinib after oral administration to humans. *Drug Metab Dispos* 2008; **36**:1357-1364.
11. He K, Lago MW, Iyer RA, Shyu WC, Humphreys WG, Christopher LJ. Lacteal secretion, fetal and maternal tissue distribution of dasatinib in rats. *Drug Metab Dispos* 2008; **36**:2564-2570.
12. Kovach JS, Moertel CG, Schutt AJ, Eagan RT. Phase I study of hycanthone. *Cancer Treat Rep* 1979; **63**:1965-1969.
13. Bodin NO, Borg KO, Johansson R, Obianwu H, Svensson R. Absorption, distribution and excretion of alprenolol in man, dog and rat. *Acta Pharmacol Toxicol (Copenh)* 1974; **35**:261-269.
14. Black JW, Duncan WA, Shanks RG. Comparison of some properties of pronethalol and propranolol. *Br J Pharmacol Chemother* 1965; **25**:577-591.
15. Walle UK, Thibodeaux H, Privitera PJ, Walle T. Stereochemistry of tissue distribution of racemic propranolol in the dog. *Chirality* 1989; **1**:192-196.
16. Dynon MK, Jarrott B, Louis WJ. Tissue distribution and hypotensive effect of prazosin in the conscious rat. *J Cardiovasc Pharmacol* 1983; **5**:235-239.
17. Berg G, Jonsson KA, Hammar M, Norlander B. Variable bioavailability of papaverine. *Pharmacol Toxicol* 1988; **62**:308-310.
18. American Regent IS, NY 11967. Prescribing information for papverine hydrochloride. 2009.
19. Michiels M, Van Peer A, Woestenborghs R, Heykants J. Pharmacokinetics and tissue distribution of astemizole in the dog. *Drug Development Research* 1986; **8**:53-62.

20. Sharma A, Hamelin BA. Classic histamine H1 receptor antagonists: a critical review of their metabolic and pharmacokinetic fate from a bird's eye view. *Curr Drug Metab* 2003; **4**:105-129.
21. Wilton J, Kurenova E, Pitzonka L, *et al.* Pharmacokinetic analysis of the FAK scaffold inhibitor C4 in dogs. *Eur J Drug Metab Pharmacokinet* 2016; **41**:55-67.
22. Thudium KE, Telang U, Wang P, *et al.* Abstract 4257: Pharmacokinetics (PK) and tissue penetration of the novel VEGFR-3/FAK inhibitor, chloropyramine. 102nd Annual Meeting of the American Association for Cancer Research; 2011; Orlando, Florida.
23. Xin-Guo Z, Kou F, Guo-Di M, Tang P, Zhong-Duo Y. Pharmacokinetics and pharmacodynamics of a novel Acetylcholinesterase Inhibitor, DMNG-3. *Acta Neurobiol Exp (Wars)* 2016; **76**:117-124.
24. Pendopharm DoP, Inc. Montreal, Quebec, Canada. Prescribing Information Bntropine Mesylate Tablets, USP 1mg BM Oral Solution 0.4mg/ml. 2015
25. Scharman EJ, Hutzler JM, Rosencrance JG, Tracy TS. Single dose pharmacokinetics of syrup of ipecac. *Ther Drug Monit* 2000; **22**:566-573.
26. Mukhopadhyay R, Roy S, Venkatadri R, *et al.* Efficacy and Mechanism of Action of Low Dose Emetine against Human Cytomegalovirus. *PLoS Pathog* 2016; **12**:e1005717.
27. Ren L, Zhao H, Chen Z. Study on pharmacokinetic and tissue distribution of lycorine in mice plasma and tissues by liquid chromatography-mass spectrometry. *Talanta* 2014; **119**:401-406.
28. Yang G, Zhang C, Hu P, Zhu M, Hu M, Gao S. An UPLC-MS/MS method for quantifying tetrandrine and its metabolite berbamine in human blood: Application to a human

- pharmacokinetic study. *J Chromatogr B Analyt Technol Biomed Life Sci* 2017; **1070**:92-96.
29. Ju A, Kang K, Xue Q, Li Q. Pharmacokinetics, tissue distribution and excretion study of tetrandrine in rats. *J Chem Pharm Sci* 2020; **24**:557-562
30. Ekins S, Lingerfelt MA, Comer JE, *et al.* Efficacy of Tilorone Dihydrochloride against Ebola Virus Infection. *Antimicrob Agents Chemother* 2018; **62**:e01711-e01717
31. Wacker A, Lodemann E, Gaur V, Diederich J. Distribution of 14 C-tilorone in mice. *Naturwissenschaften* 1972; **59**:520.
32. Knudsen JB, Bastain W, Sefton CM, Allen JG, Dickinson JP. Pharmacokinetics of ticlopidine during chronic oral administration to healthy volunteers and its effects on antipyrine pharmacokinetics. *Xenobiotica* 1992; **22**:579-589.
33. Smith TC, Kinkel AW, Gryczko CM, Goulet JR. Absorption of pyrvinium pamoate. *Clin Pharmacol Ther* 1976; **19**:802-806.
34. Swart KJ, Sutherland FC, van Essen GH, Hundt HK, Hundt AF. Determination of fluspirilene in human plasma by liquid chromatography-tandem mass spectrometry with electrospray ionisation. *J Chromatogr A* 1998; **828**:219-227.
35. Sklerov J, Levine B, Moore KA, Allan C, Fowler D. Tissue distribution of loperamide and N-desmethyloperamide following a fatal overdose. *J Anal Toxicol* 2005; **29**:750-754.
36. Shang E, Xiang B, Liu G, *et al.* Determination of phenazopyridine in human plasma via LC-MS and subsequent development of a pharmacokinetic model. *Anal Bioanal Chem* 2005; **382**:216-22.
37. Thomas BH, Whitehouse LW, Solomonraj G, Paul CJ. Metabolism and disposition of phenazopyridine in rat. *Xenobiotica* 1993; **23**:99-105.

38. Gao X, Zhang Y, Wang Y, *et al.* Influence of verapamil on pharmacokinetics of pristimerin in rats. *Biomed Chromatogr* 2016; **30**:802-809.
39. Burns JJ, Yu TF, Berger L, Gutman AB. Zoxazolamine: physiological disposition, uricosuric properties. *Am J Med* 1958; **25**:401-408.
40. Weston S, Haupt R, Logue J, Matthews K, Frieman M. FDA approved drugs with broad anti-coronaviral activity inhibit SARS-CoV-2 *in vitro*. *bioRxiv* 2020; 10.1101/2020.03.25.008482
41. He Y, Pei R, Xu Z, *et al.* Mycophenolate mofetil is active against SARS-CoV-2 in Vero E6 cells. *Preprints* 2020; 202004.0380.v1
42. Ianevski A, Yao R, Fenstad M, *et al.* Potential Antiviral Options against SARS-CoV-2 Infection. *Viruses* 2020; **12**:642
43. Zhang Y, Zhang Q, Li X, *et al.* Gemcitabine, lycorine and oxysophoridine inhibit novel coronavirus (SARS-CoV-2) in cell culture. *Emerging Microbes and Infections* 2020; **9**:1170-1173
44. Ellinger B, Bojkova D, Zaliani A, *et al.* Identification of inhibitors of SARS-CoV-2 *in vitro* cellular toxicity in human (Caco-2) cells using a large scale drug repurposing collection. *Research Square* 2020; 10.21203/rs.3.rs-23951/v1